

Cholesterol in brain disease: sometimes determinant and frequently implicated

Mauricio G Martín^{1,*}, Frank Pfrieger^{2,**} & Carlos G Dotti^{3,***}

Abstract

Cholesterol is essential for neuronal physiology, both during development and in the adult life: as a major component of cell membranes and precursor of steroid hormones, it contributes to the regulation of ion permeability, cell shape, cell–cell interaction, and transmembrane signaling. Consistently, hereditary diseases with mutations in cholesterol-related genes result in impaired brain function during early life. In addition, defects in brain cholesterol metabolism may contribute to neurological syndromes, such as Alzheimer's disease (AD), Huntington's disease (HD), and Parkinson's disease (PD), and even to the cognitive deficits typical of the old age. In these cases, brain cholesterol defects may be secondary to disease-causing elements and contribute to the functional deficits by altering synaptic functions. In the first part of this review, we will describe hereditary and non-hereditary causes of cholesterol dyshomeostasis and the relationship to brain diseases. In the second part, we will focus on the mechanisms by which perturbation of cholesterol metabolism can affect synaptic function.

Keywords brain disease; cholesterol metabolism; cognition

DOI 10.15252/embr.201439225 | Received 26 June 2014 | Revised 13 August 2014 | Accepted 18 August 2014 | Published online 15 September 2014

EMBO Reports (2014) 15: 1036–1052

See the Glossary for abbreviations used in this article.

Introduction

Cholesterol is an essential constituent of eukaryotic membranes, and as such, it impacts on nearly all aspects of cellular structure and function [1–5]. In addition, cholesterol serves as precursor for steroid hormone and bile acid synthesis [6] and therefore has a critical role in body metabolism (Fig 1). Furthermore, cholesterol can also influence cell function through its biologically active oxidized forms: oxysterols [7–11].

Cellular cholesterol synthesis is a complex and resource-intensive process: it starts with the conversion of acetyl-CoA to 3-hydroxy-3-methylglutaryl-CoA by HMG-CoA synthase, which is then converted

to mevalonate by HMG-CoA reductase. The latter constitutes the only known rate-limiting and irreversible step in cholesterol synthesis. This is followed by a long series of enzymatic reactions that convert mevalonate into 3-isopentenyl pyrophosphate, farnesyl pyrophosphate, squalene, lanosterol and, through a 19-step process, to cholesterol (Fig 1) [12]. Whether other enzymatic steps in the pathway, posterior to squalene synthesis, are rate-limiting is unknown. Cholesterol is synthesized in the endoplasmic reticulum from where it is distributed to cellular membrane compartments by vesicular and non-vesicular transport mechanisms (Fig 1) [13]. Cells can also import sterols through receptor-mediated endocytosis of lipoproteins and the subsequent export of unesterified cholesterol from lysosomes (Fig 1) [14,15]. It is well established that cells regulate their cholesterol content by an exquisite feedback mechanism that balances biosynthesis and import. Cells sense their level of cholesterol by membrane-bound transcription factors known as sterol regulatory element-binding proteins (SREBPs), which regulate the transcription of genes encoding enzymes of cholesterol and fatty acid biosynthesis as well as lipoprotein receptors [16].

Cholesterol is not uniformly distributed within membranes and across different cellular compartments. A recent study *in vitro* suggests that cholesterol is enriched in the cytosolic (inner) leaflet of the plasma membrane [17]. Moreover, there is evidence that the plasma membrane contains nano/micro-domains that are enriched in cholesterol, the so-called 'lipid rafts'. These rafts are thought to represent highly dynamic structures dispersed throughout the membrane of cells that recruit downstream signaling molecules upon activation by external or internal signals [18]. Rafts also contain a high amount of sphingomyelin, which is enriched in the outer leaflet of the plasma membrane, indicating that some trans-bilayer translocation must occur to form and stabilize these domains [19]. In neurons, membrane rafts have been detected at synapses, where they are thought to contribute to pre- and postsynaptic function [20–24].

The brain contains 23% of all cholesterol in the body [25], and within the brain, a large fraction of cholesterol is present in the myelin sheath that is formed by oligodendrocytes to insulate axons (Fig 2). Neurons and astrocytes probably also contain large amounts of cholesterol to maintain their complex morphology and synaptic transmission. All cells in the brain are cut off from

1 Instituto de Investigaciones Médicas Mercedes y Martín Ferreyra (INIMEC-CONICET-UNC), Universidad Nacional de Córdoba, Córdoba, Argentina

2 Institute of Cellular and Integrative Neurosciences, CNRS UPR 3212, University of Strasbourg, Strasbourg, France

3 Centro Biología Molecular 'Severo Ochoa' CSIC-UAM, Madrid, Spain

*Corresponding author. Tel: +54 351 468 1464; Fax: +54 351 469 5163; E-mail: mmartin@imfm.uncor.edu

**Corresponding author. Tel: +3 338 845 6645; Fax: +3 338 860 1664; E-mail: frank.pfrieger@inci-cnrs.unistra.fr

***Corresponding author. Tel: +34 91 196 4519; Fax: +34 91 196 4420; E-mail: cdotti@cbm.csic.es

Glossary

24-OHC	24S-hydroxycholesterol
ABCA1	ATP-binding cassette transporter A1
AD	Alzheimer's disease
AKAP150	A-kinase-anchoring protein 150
Akt	serine/threonine kinase Akt/PKB (Protein kinase B)
AMPARs	α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptors
APOE	apolipoprotein E
APP	amyloid precursor protein
CLU	Clusterin
CNS	central nervous system
Cox-2	cyclooxygenase 2
CSF	cerebrospinal fluid
CYP46	cytochrome P450 cholesterol 24-hydroxylase
CYP7B1	cytochrome P450 oxysterol 7- α -hydroxylase
dhcr24	24-dehydrocholesterol reductase
eNOS	endothelial nitric oxide synthase
EPSC	excitatory postsynaptic currents
GWAS	genome-wide association studies
HD	Huntington's disease
HDL	high density lipoprotein
HMG-CoA	3-hydroxy-3-methylglutaryl-CoA
LDL	low density lipoprotein
LRRK2	leucin-rich repeat-kinase 2
LTD	long-term depression
LTP	long-term potentiation
LXRs	liver X receptors
MARCKS	myristoylated alanine-rich C kinase substrate
Mdm2	mouse double minute 2 homolog
Mecp2	methyl CpG binding protein 2
MβCD	methyl- β -cyclodextrin
NMDAR	N-methyl-D-aspartate receptor
NPC	Niemann-Pick C
PD	Parkinson's disease
PI(4,5)P2	phosphatidylinositol-(4,5)-bisphosphate
PINK1	PTEN-induced putative kinase 1
PLCγ	phospholipase C γ
PSD95	postsynaptic density protein 95
PUFAs	polyunsaturated fatty acids
ROS	reactive oxygen species
SLOS	Smith–Lemli–Opitz syndrome
SNAREs	Soluble NSF Attachment Protein REceptors
SREBPs	sterol regulatory element-binding proteins
SV	synaptic vesicle
T2DM	comorbid type 2 diabetes mellitus
TrkB	tropomyosin-related kinase B receptor

cholesterol supply by the blood, because the blood–brain barrier prevents entry of cholesterol-rich lipoproteins. Therefore, all cholesterol in the CNS is made locally [26,27]. The fact that brain cholesterol metabolism is separated from the rest of the body warrants caution when causal correlations between high blood cholesterol and brain pathologies are suggested: in the presence of an intact blood–brain barrier neurons and all other cells in the brain are not influenced by circulating cholesterol (Fig 2). On the other hand, there is a constant efflux of cholesterol from the brain, which is enabled by the neuron-specific cytochrome P450 oxidase Cyp46a1. This enzyme hydroxylates cholesterol to 24S-hydroxycholesterol (24-OHC), which crosses the blood–brain barrier, enters the circulation and is metabolized by the liver (Fig 2) [28].

In the mouse brain, cholesterol synthesis is high until the second post-natal week and then decreases significantly and remains low

in the adult [29]. Age-associated cholesterol reduction was also observed in postmortem human samples [30]. Importantly, the rate of cholesterol synthesis in individual brain cells is unknown. Whereas oligodendrocytes probably rely on their own cholesterol synthesis to form and maintain myelin, neurons may import cholesterol from astrocytes. The high metabolic rate of neurons probably enforces a constant turnover of cholesterol throughout life (Fig 2) [31].

Enzymatic deficiencies in the cholesterol synthesis pathway are responsible for a number of inherited disorders with severe neurodevelopmental defects. In addition, alterations in brain cholesterol are thought to be critically involved in a number of neurodegenerative pathologies, in some cases as the suspected cause and in others as a consequence. In the following sections, we will summarize our current knowledge of how defects in cholesterol metabolism and transport can lead to, or be part of, brain dysfunction.

Cholesterol in brain disease: hereditary causes

The important role of cholesterol for CNS function is exemplified by a number of rare hereditary diseases caused by mutations in cholesterol-related genes. The reader interested in a comprehensive description of these diseases should refer to excellent reviews [32–34]. Here, we will focus on three diseases that are linked to mutations in cholesterol metabolism and transport steps: the Smith–Lemli–Opitz syndrome, desmosterolosis and the Niemann–Pick type C disease. In addition, we will briefly comment on the recent discovery of an association between cholesterol metabolism and Rett syndrome.

Smith–Lemli–Opitz syndrome (SLOS) is a rare autosomal recessive disorder caused by mutations in the gene encoding the enzyme 7-dehydrocholesterol reductase (DHCR7, Fig 1). This enzyme catalyzes the last step in the Kandutsch–Russell pathway of cholesterol biosynthesis. There are reports of 154 different mutations in DHCR7 in SLOS patients [35]. In the most severe cases, the mutation causes fetal or newborn death, due to severe malformations and multiple organ failure. The milder cases can present multi organ defects of different severity: facial and cranial malformations, hypospadias or complete gonadal absence and even gender reversal, gastrointestinal symptoms, limb malformations, liver disease and cardiac defects (for comprehensive descriptions see [36–38]). SLOS patients present, with different degree of gravity, intellectual disabilities, delayed motor and language maturation, affective disorders and sleeping problems. The signs and symptoms may be due to reduced brain cholesterol levels. Severely affected SLOS patients have plasma cholesterol concentrations at 2% of the normal level and low cholesterol content in all tissues, especially in the brain [39]. Patients with milder symptoms can show normal plasma concentrations of cholesterol, probably due to residual synthesis capacity and dietary import. However, even in these mild cases, circulating cholesterol cannot compensate for the brain deficits due to the blood–brain barrier, suggesting that low cholesterol levels in the brain are the primary cause of the neurological symptoms in SLOS patients. Alternatively, these symptoms may be due to accumulation of 7,8-dehydrodesmosterol, which is the substrate of DHCR7, or its oxidized metabolites [36]. Notably, some of the teratogenic effects by impaired cholesterol synthesis are probably due to defects in

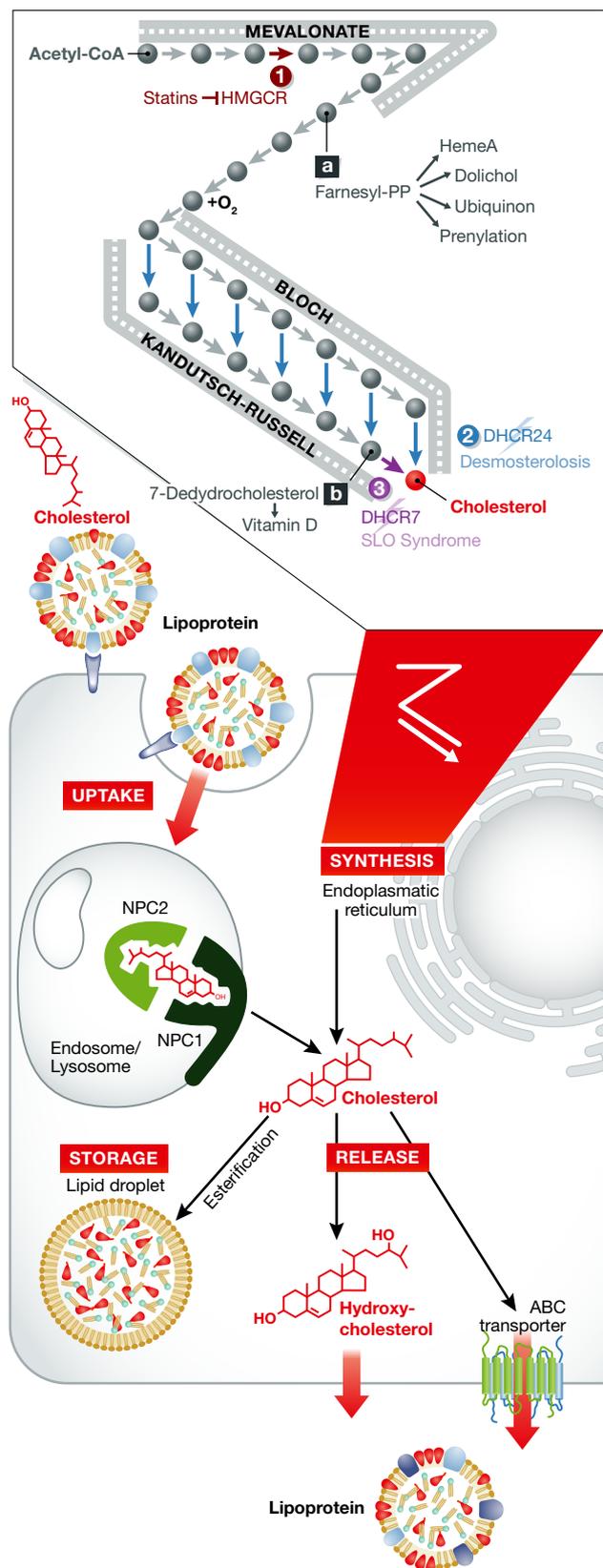


Figure 1. Cellular cholesterol homeostasis.

Diagram summarizing how cells ensure cholesterol homeostasis. Cells synthesize cholesterol from acetyl-CoA by a long series of enzymatic steps requiring energy and molecular oxygen. Intermediates of the pathway serve as precursors for other biologically active molecules. Highlighted enzymes are 3-hydroxy-3-methyl-glutaryl-CoA reductase (HMGCR), which is rate-limiting for the mevalonate pathway and inhibited by statins, 24-dehydrocholesterol reductase (DHCR24) and 7-dehydrocholesterol reductase (DHCR7), whose defects cause rare human diseases. Cells can take up cholesterol by receptor-mediated endocytosis of lipoproteins bearing apolipoproteins. In this pathway, Niemann-Pick Type C Protein 1 and NPC2 mediate cooperatively the exit of cholesterol out of the endosomal-lysosomal system and thereby allow for its incorporation into the intracellular pool. Defects in either protein cause the lysosomal storage disorder Niemann-Pick Type C. Overload by cholesterol is prevented by its intracellular esterification and subsequent storage in lipid droplets and by its release. Cholesterol is released either as a complex with apolipoprotein-containing lipoproteins via members of the ATP-binding cassette transporters or after conversion to oxysterols. Exemplary proteins for each process are indicated. The relative contribution of each pathway to cholesterol homeostasis is probably cell type-specific. The post-lanosterol steps of cholesterol biosynthesis are divided into Bloch and Kandutsch-Russell pathways, which share enzymatic stages but produce C24 double-bond reduced cholesterol at different steps.

Desmosterolosis is a rare, autosomal recessive disease caused by mutations in the *dhcr24* gene (Fig 1). The enzyme encoded by this gene catalyzes the reduction of desmosterol to cholesterol, the last step in the Bloch pathway of cholesterol synthesis [44]. *dhcr24* was first identified as a gene involved in human steroid synthesis due to its similarity to DIMINUTO/DWARF1 (*dwf1*). The product of this gene catalyzes a similar step in sterol synthesis in plants, which is essential for normal growth and development in *Arabidopsis* [45,46]. In addition to its enzymatic activity, DHCR24 contains a binding site for p53 and Mdm2, which mediate oncogenic and oxidative stress signaling [47]. Patients with DHCR24 mutations present severe brain defects including microcephalia, hydrocephalia, ventricular enlargement, defects in the corpus callosum, and thinning of white matter and seizures. Disease signs may be attributed to reduced cholesterol levels or to accumulation of desmosterol. In addition, reduced levels of DHCR24 in the adult brain may enhance sensitivity to oxidative stress [48,49].

Niemann-Pick C disease is an autosomal recessive disorder caused in 95% of the cases by mutations in the *npc1* or *npc2* genes, which cause progressive visceral, neurological, and psychiatric symptoms and premature death. Neurologic symptoms and the age of onset vary strongly among patients. The symptoms include delayed development of motor skills, supranuclear palsy, gait problems, frequent falls, clumsiness, difficulties to speak and learn, and ataxia. The adult form presents psychiatric problems, cerebellar ataxia, and progressive dementia. Cataplexy, seizures, and dystonia are other common features [50,51]. At the histological level, NPC-deficient brains present neurons with enlarged neurites, ectopic dendrites, neurofibrillary tangles, neuroinflammation, and axonal dystrophy. In the advanced disease, neuronal death is the prominent feature, affecting specific cell types, particularly cerebellar Purkinje cells [52]. At the cellular level, NPC1 and NPC2 cooperate to mediate the exit of cholesterol from the endosomal-lysosomal system (Fig 1) [53–55]. Defects in either protein cause accumulation of unesterified cholesterol and gangliosides in late endosomes and lysosomes [56–58] with reduced cholesterol levels in the plasma membrane of NPC patient cells [59] and in axons of cultured neurons [60]. Moreover, endosomal organelle transport [61] and

sonic hedgehog signaling [40,41]. During autoprocessing of sonic hedgehog, a cholesterol molecule is covalently added to the amino terminus and regulates protein function [42,43].

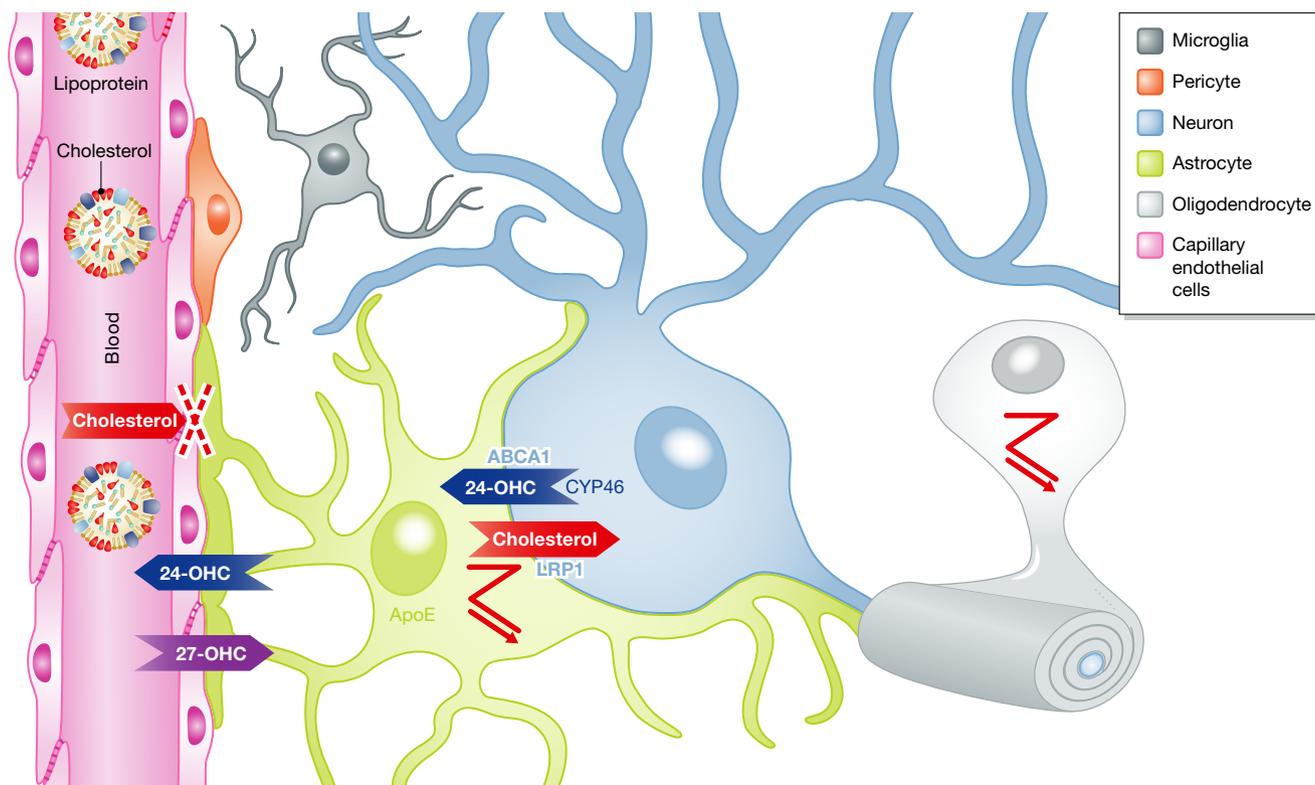


Figure 2. Brain-specific aspects of cholesterol metabolism.

Brain cells are cut off from blood supply as the blood–brain barrier prevents entry of lipoproteins. Cholesterol is synthesized by different types of glial cells as indicated by the zagged pathway symbol. Some neurons may import cholesterol from APOE-expressing astrocytes via lipoprotein uptake (LRP1) and hydroxylate surplus cholesterol to 24-OHC, which is excreted via ABCA1 and which enters the blood circulation. 24-OHC is able to cross the blood–brain barrier where it may signal the level of plasma cholesterol levels to cells in the brain. Cholesterol-related proteins with cell-specific distribution are indicated.

synaptic vesicle composition and morphology are affected [62]. Hence, the neurological and psychiatric symptoms in NPC disease may result, like in the previously mentioned diseases, from a combination of abnormal lipid accumulation and reduced cholesterol levels in specific membrane compartments [63].

Rett syndrome is an X-linked neurological disorder characterized by defective motor control, cognitive abilities, and social interactions, namely appearance of speech difficulties, stereotypic hand movements, problematic walking, seizures, intellectual disabilities, and autistic behavior. Rett syndrome is caused by mutations in the X-linked *mecp2* gene, which encodes a methyl DNA-binding protein that regulates gene expression [64]. Until recently, Rett syndrome was not associated with a defect in cholesterol metabolism. However, a recent study has shown that a suppressing mutation in the *sqle* gene, encoding for the squalene epoxidase that mediates a dedicated step in cholesterol synthesis, was sufficient to restore function and longevity in *Mecp2* null mice [65]. These data suggest that *Mecp2* may be involved in the transcriptional control of cholesterol metabolism. In support of this hypothesis, the expression of 3-hydroxy-3-methylglutaryl-coA reductase, squalene epoxidase, and Cyp46A1 was altered in *Mecp2* null mice in an age-dependent manner. Measurements of de novo cholesterol synthesis confirmed that sterol synthesis decreases in adult brains of *Mecp2* mutant mice. Administration of fluvastatin, an HMG-CoA reductase inhibitor with intermediate capacity to cross the blood–brain barrier [66], to

Mecp2 null mice lowered serum cholesterol levels, improved motor, cognitive and social behaviors and increased life span. In summary, while Rett syndrome is clearly associated with changes in cholesterol metabolism, it remains to be demonstrated whether *Mecp2* phenotypes are caused by neuron-specific defects in cholesterol metabolism [67].

Taken together, these rare genetic diseases underline that the brain is highly sensitive to perturbations of cholesterol synthesis and transport and that these defects cannot be rescued by manipulations of peripheral cholesterol levels.

Brain cholesterol and Alzheimer's disease

Apart from hereditary diseases with a clear-cut link to cholesterol, there are neurodegenerative diseases for which a contribution of impaired cholesterol metabolism to the disease has been proposed. One of the most studied cases is Alzheimer's disease (AD).

Twenty years ago, it was shown that the apolipoprotein E allele $\epsilon 4$ enhances the risk of late-onset familial and sporadic AD [68] and increases amyloid beta deposition in the brain [69]. This ApoE variant is also associated with cardiovascular disease, atherogenicity, and high LDL-cholesterol levels in blood [70–72], raising the possibility that peripheral cholesterol dyshomeostasis could contribute

to late-onset AD. Consistently, early work showed that rabbits fed with a high cholesterol diet develop intracellular accumulations of beta amyloid in brain cells [73]. Later on, clinical retrospective studies in humans showed that hypercholesterolemia predisposes to cognitive deficits, including dementias of the Alzheimer's type [74], and that chronic treatment with cholesterol-lowering statins seemed to prevent the disease [75,76]. More recently, a multisite, medical center-based analysis of early patients with AD confirmed a correlation between high LDL-cholesterol, low HDL-cholesterol and high PIB index, which measures the levels of cerebral A β with carbon C11-labeled Pittsburgh Compound B [77]. Several lines of evidence suggest that the link between elevated blood cholesterol and AD is related to vascular and inflammatory alterations (and associated diseases) rather than through changes in brain cholesterol [78–88]. In fact, atherosclerosis affects the cerebral vasculature, leading to plasma protein leakage, accumulation of lipid-containing macrophages, and vessel fibrosis, which causes hypoperfusion, local inflammation and, eventually, breakdown of the blood–brain barrier [78–80]. Moreover, there is good evidence that metabolic conditions characterized by high circulating cholesterol levels predispose to AD through gradual and persistent vascular defects in the brain [81–86]. In agreement with the clinical evidence, chronic hypoperfusion of the brain in rats and AD mice increased BACE1 expression, the concentration of A β fibrils and caused cognitive impairment [87,88]. In addition to hypoperfusion and inflammation, high blood cholesterol may also cause vascular defects predisposing to AD via oxysterols. These metabolites contribute to the atherogenic process by inducing endothelial cell dysfunction, adhesion of circulating blood cells, foam cell formation, and apoptosis of vascular cells [89]. Furthermore, hypercholesterolemia may predispose to AD by comorbid type 2 diabetes mellitus. Clinical studies revealed that a high proportion of patients with AD present T2DM [90], which often associates with hypercholesterolemia [91]. In this constellation, AD signs may also result from defects in brain insulin signaling [92].

On the other hand, predisposition to AD in cases of familial hypercholesterolemia with the $\epsilon 4$ variant of ApoE may result from isoform-specific effects on brain cells. This could involve alterations in the metabolism of beta peptide, effects on synaptic function, and disturbance of cholesterol metabolism in the CNS [93]. Animal studies have shown that ApoE levels are reduced in the brain of mice expressing human Apo $\epsilon 4$, suggesting a shorter life span of this isoform. Moreover, Apo $\epsilon 4$ has different properties than other Apo ϵ isoforms, which may impair the glia-to-neuron transport of cholesterol, reduce myelin, and diminish the capacity to degrade the toxic amyloid peptide [94–100]. Still, the consequences of the inheritance of the $\epsilon 4$ allele ought to be put together with the vascular defects due to hypercholesterolemia and with the direct toxic effects of $\epsilon 4$ on the cerebrovascular system [101]. Interestingly, a recent study on a cohort of NPC patients revealed a correlation between ApoE alleles and disease severity, which confirms an impact of cholesterol dyshomeostasis on neurodegeneration.

Recent data from genome-wide association studies (GWAS) suggest that mutations in the lipid metabolism-related proteins ApoJ/Clusterin (CLU) and ABCA7 are risk factors for AD [102]. ApoJ/Clusterin is involved in lipid transport but also has heat-shock-like chaperone activity and regulates apoptosis, immunoglobulin interaction and complement defense [103,104]. Individuals

carrying particular polymorphisms present elevated levels of LDL-cholesterol and vascular defects [105]. Aging mice deficient in ApoJ/Clusterin develop progressive glomerulopathy characterized by deposition of immune complexes in the thin membranes surrounding the capillary loops [106]. Mutations or polymorphisms in the ABCA family member ABCA1 are associated with Tangier disease or, in the less severe cases, with familial HDL deficiency. Both conditions are characterized by reduced levels of circulating HDLs and the deposition of cholesterol esters in peripheral tissues [107]. Hence, like for the $\epsilon 4$ allele of ApoE, polymorphisms in these two proteins may predispose to AD by peripheral, vascular defects with or without direct changes in brain cholesterol. Future research will need to address what the relative and specific contributions of the diverse defects to AD predisposition are.

A link between brain cholesterol and AD was also suggested by several *in vitro* studies on neuronal, pseudo-neuronal, and non-neuronal cells over-expressing proteins involved in familial AD. Acute reduction of cholesterol levels after treatment of these cells with synthesis inhibitors like statins or sterol-extracting drugs as methyl- β -cyclodextrin, reduced the production and toxicity of amyloid beta peptide, suggesting that elevated levels of brain cholesterol can be the cause of AD [108]. Although early work showed that brain cholesterol is high in the brains of patients with AD, as a consequence of excess A β [109], others showed reduced total brain cholesterol and brain cholesterol synthesis in Alzheimer's disease patients [110–117]. These studies reported a reduction in the lipid bilayer width of neurons, a 30% reduction in free, unesterified cholesterol in the temporal gyrus but not in the cerebellum [113] and a significant reduction of cholesterol levels in the whole-brain raft fraction [116] and in the white matter [114]. On the other hand, cholesterol levels were increased in nerve terminals of AD brains that were rich in amyloid aggregates [118] and in the core of mature, but not diffuse, amyloid plaques [119]. These observations suggest that amyloid-mediated sequestration of membranes, mainly in nerve terminals, may be one of the causes for cholesterol depletion from neuronal membranes in AD brains. Other causes of low cholesterol levels in AD brains may be APP-mediated inhibition of cholesterol synthesis [120], amyloid beta peptide-induced reduction of ApoE-mediated cholesterol uptake [121], increased CYP46 activity leading to cholesterol oxidation and excretion [122,123] or beta amyloid-induced modifications of lipid rafts [124]. Increased Cyp46 activity leading to cholesterol elimination could possibly be one of the multiple consequences of amyloid peptide-induced synaptic calcium alterations and oxidative stress [125,126]. These different possibilities are illustrated in Figure 3. In agreement with the observation that AD is accompanied by low brain cholesterol, low levels of cholesterol are also characteristic of the aging human brain in regions susceptible to the disease like the hippocampus [30,127–129], in Alzheimer's-like mice [130] and in hippocampal synapses of old mice [131]. *In vitro* studies have proposed that low neuronal membrane cholesterol levels might contribute to AD by a combination of events, including an increase in beta amyloid peptide production [132], a reduction in beta amyloid peptide degradation [133], an increase in the inflammatory response [134] and by facilitating the interaction of A β 42 oligomers with lipid rafts, leading to plasma membrane perturbation, calcium dyshomeostasis, and toxicity [124,135]. In fibroblasts from Alzheimer's patients, recruitment of amyloid assemblies to the plasma membrane is higher in cholesterol

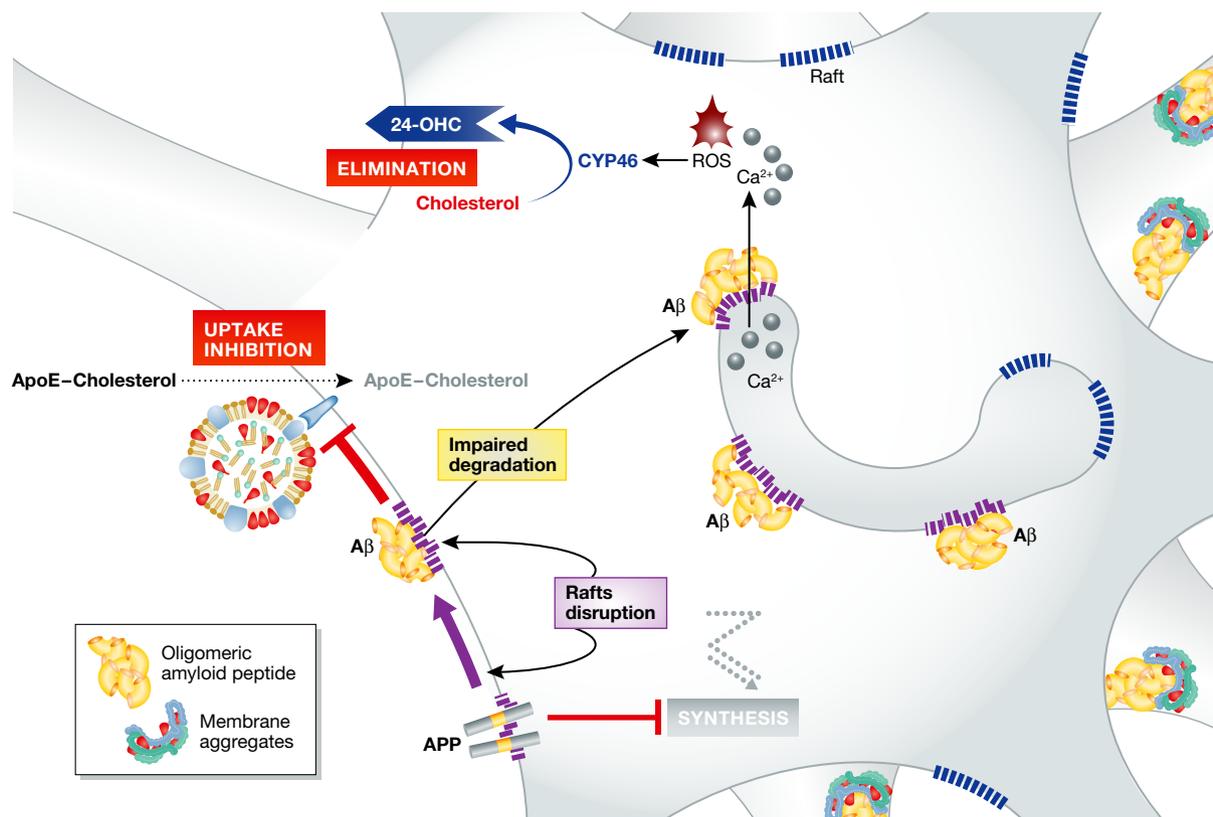


Figure 3. Schematic picture showing how AD can lead to reduced cholesterol in neurons.

Numerous studies in brain samples from AD affected individuals show reduced cholesterol levels in structures like the hippocampus. *In vitro* studies suggest that amyloid oligomers, whether through binding to detergent resistant membrane domains (rafts) and/or in the form of amyloid peptide aggregates in intracellular compartments, trigger cell cholesterol decrease by various mechanisms: stress-activated Cyp46A1 transcription leading to cholesterol solubilization and excretion in the form of 24-OHC; sequestration of membranes and cholesterol within terminals from dying neurons and directly in amyloid plaques; inhibition of astrocyte-derived ApoE-cholesterol uptake and/or by direct changes in plasma membrane lipid content. Cholesterol reduction can also occur in conditions accompanied by an excess of APP through direct inhibition of HMG-CoA reductase (HMGCR) and SREBP mRNA levels.

poor conditions while high membrane cholesterol levels in these cells prevent A β 42-induced generation of ROS and membrane lipo-peroxidation [136].

In summary, while it is quite clear that pharmacological reduction of cholesterol levels in cells with normal cholesterol content can inhibit amyloidogenic processing of over-expressed APP in a variety of cell types and systems [reviewed in 108], it remains unclear whether the conclusions from these studies can be extrapolated to the disease or normal aging contexts, especially taking into account the human studies showing that cholesterol levels are reduced in normal and pathological (Alzheimer's) aging brains (see Sidebar A). Given the numerous functions of cholesterol, it is not difficult to envision how reduced neuronal cholesterol levels can lead to brain dysfunction.

Brain cholesterol metabolism is altered by neurodegenerative pathologies and during aging

Significant changes in brain cholesterol metabolism have also been observed in other pathological conditions different from AD, such as Huntington's disease (HD), Parkinson's disease (PD), depression, amyotrophic lateral sclerosis, stroke, head trauma, and also normal aging.

Aging is characterized by cognitive decline and is accompanied by altered short-term memory and learning. An age-dependent loss of cholesterol has been observed in the human brain [30,112,127–129] and in the rodent hippocampus, *in vivo* and *in vitro* [131,137]. At present, the causes for the age-associated cholesterol loss are not known. There are several candidate mechanisms. First, it could be due to increased transcriptional activation [125] and membrane mobilization [138] of the brain-specific cholesterol-hydroxylating enzyme Cyp46A1. This is supported by the observations that Cyp46A1 increases in high stress situations, such as cortical injury, induced autoimmune encephalomyelitis and in Alzheimer's disease [123,139,140]. Alternatively, the age-dependent lowering of cholesterol levels may be due to reduced synthesis in neurons or impaired delivery from glial cells.

Huntington's disease (HD) is an autosomal dominant neurodegenerative disorder caused by an abnormal expansion of a CAG repeat in the huntingtin gene resulting in behavioral abnormalities, cognitive decline, and involuntary movements. In HD patients, cholesterol metabolism in the brain is impaired [141,142] possibly by inhibition of SREBPs [141] by mutant huntingtin. Valenza and colleagues showed that cholesterol biosynthesis is reduced in brain samples from different transgenic mouse models expressing mutant huntingtin [143]. The authors also showed that cholesterol is first

reduced in synaptosomes and later on in myelin and that transcript levels of genes mediating cholesterol biosynthesis and efflux are reduced in HD astrocytes causing lower production and secretion of ApoE from these cells. Notably, these results are in contradiction with other studies showing increased free cholesterol in the same mice [144]. The divergence may be due to the use of different experimental approaches, namely mass spectrometry by Valenza *et al* [143] and filipin staining and thin-layer chromatography by Trushina *et al* [144]. Moreover, Valenza *et al* [141] perfused animals with saline before measurements of sterol levels in the brain, which may have reduced a substantial fraction of blood-derived cholesterol. Further support that mutant huntingtin affects cholesterol levels comes from *in vitro* experiments showing that the biosynthesis of cholesterol and fatty acids is impaired in cells expressing disease-causing mutants of huntingtin [141]. HD patients and HD mice show a progressive decrease of 24-OHC [142,143], which is a sign of neuronal cell loss. The decrease in this metabolite may further reduce cholesterol levels in the brain by impairing the activity of liver X receptors (LXRs), thus reducing expression of LXR-dependent transcripts like ABCA1 and ApoE and consequently the transport of cholesterol to neuronal cells. Evidence for a role of the LXR pathway in HD—but not necessarily in ABCA1, ApoE and cholesterol levels in the brain—comes from the observation that treatment with a LXR agonist partially reverts symptoms in a zebrafish model of HD [145].

Parkinson's disease (PD) is the second most common progressive neurodegenerative disorder after AD. Like in AD, a low number of PD cases are caused by mutations in specific genes: α -synuclein, parkin, LRRK2, PINK1, DJ-1, and ATP13A2 [146]. PD is characterized by the degeneration of dopaminergic neurons in the substantia nigra of the midbrain and by the development of Lewy Bodies in neurons. Postmortem studies of highly purified lipid rafts from the frontal cortex of control, early motor stages PD, and incidental PD subjects did not reveal disease-related differences in the contents of sphingomyelin or cholesterol. However, the levels of polyunsaturated fatty acids were significantly reduced in raft fractions from PD compared to age-matched control subjects. Significant reductions were also observed in the fatty acid 18:1 (n-9) in combination with significant increases in stearic acid (18:0). The authors proposed that these changes could determine how cells respond to different forms of physical and/or chemical stress. However, confounding differences in dietary lipid uptake between patients and control subjects must be taken into account. On the other hand, acute manipulations of cholesterol *in vitro* suggest that cholesterol defects are not only a consequence of PD pathology but could contribute to PD. Treatment of cells and mice with the cholesterol-extracting drug methyl- β -cyclodextrin decreases the levels of α -synuclein in membrane fractions and reduces the accumulation of α -synuclein in the neuronal cell body and in synapses, preventing its aggregation [147]. However, the effects observed with cyclodextrins cannot be used to strongly argue that high cholesterol levels cause α -synuclein accumulation. First of all, it is unclear whether synuclein aggregation occurs in a background of high cellular cholesterol, and if so in which compartments. Second, and perhaps most critical, cyclodextrin represents a rather harsh treatment that not only leads to cholesterol extraction and redistribution to different membrane organelles but, due to its chemical nature, it can bind and redistribute other hydrophobic molecules (i.e. lipids) potentially affecting

other pathways. On the other hand, also statins reduce the aggregation of α -synuclein in cultured neurons and in animal models of synucleinopathies [148,149], suggesting again that high cellular cholesterol could be at the base of this pathological sign. Yet, care should be taken when interpreting these results as (i) these data were obtained in cells and animals with normal and not high cholesterol levels, and (ii) statins play numerous HMG-CoA-independent roles, including antioxidant and anti-inflammatory roles and also impact on the nitric oxide synthase pathway [150,151].

A dysregulation of brain cholesterol has been discussed in the context of other pathologic conditions including stroke, schizophrenia, depression, and amyotrophic lateral sclerosis, but again, it remains unclear whether the changes in brain cholesterol are at the base of these pathologies or are an epiphenomenon. A clear exception is spastic paraplegia type 5, a hereditary autosomal recessive disease with neurologic symptoms caused by mutations in CYP7B1 [152]. This enzyme is expressed in the brain and in the liver, where it hydroxylates 25- and 27-hydroxycholesterol to 7 α -25-dihydroxycholesterol and 7 α -27-dihydroxycholesterol. So far, it is unclear why its dysfunction causes progressive neuropathy in humans [153]. However, the strong increase in the substrate (27-hydroxycholesterol) in the CSF has been proposed to be the cause for the neurological symptoms [154].

In summary, the studies described above indicate that cholesterol metabolism in the brain is affected by various disease conditions. A major challenge for future research will be to determine the cellular origin (neurons, astrocytes, and oligodendrocytes) of disease-related changes in cholesterol levels, how each pathological condition affects cholesterol homeostasis in the brain, and whether cholesterol dyshomeostasis plays a causal role in the disease (see Sidebar A).

Mechanisms by which cholesterol dyshomeostasis could contribute to disease: role of oxysterols

Cells in the brain rely on constant cholesterol synthesis; however, the blood–brain barrier prevents entry and exit of lipoproteins. Consequently, cells in the brain require a specific mechanism to prevent an accumulation of excess cholesterol. Different mechanisms for removal of brain cholesterol are currently recognized [25,155–157]. An important one is the hydroxylation of cholesterol to 24-OHC by cholesterol 24-hydroxylase or CYP46 [25,28,157–159]. Interestingly, CYP46 is expressed specifically by neurons, suggesting that these cells are particularly sensitive to excess of cholesterol [31]. However, knockout mice lacking CYP46 do not present increased cholesterol levels, possibly due to the concomitant reduction in the cholesterol mevalonate pathway [160]. These mice exhibit severe deficiencies in spatial, associative, and motor learning, which are reversed by treatment with geranylgeraniol, a non-sterol isoprenoid required for learning, but not cholesterol. The explanation is that brains from mice lacking CYP46 excrete cholesterol more slowly, and the tissue compensates by suppressing the mevalonate pathway, which in turn results in lower synthesis of geranylgeraniol [160]. Geranylgeraniol is posttranslationally attached to a large number of proteins and regulates multiple cellular processes ranging from intracellular signaling to vesicular transport [161].

Cholesterol turnover catalyzed by CYP46 seems to be essential for neural function. Female homozygous transgenic mice that ubiquitously overexpress the *cyp46A1* gene under the β -actin promoter present levels of circulating 24-OHC that are 30–60% higher than heterozygote littermates and show increased expression of synaptic proteins and improved spatial memory in the Morris water maze test, compared to wild-type mice [162]. Curiously, this effect was not observed in male mice, implying hormonal-dependent sensitivity to the effects of 24-OHC. It is unclear whether neuronal or glial cholesterol levels contribute to the improved cognition in these mice, as cholesterol levels were not measured in this model. Beneficial effects for CYP46 have also been proposed based on the evidence that increased *cyp46A1* expression in the brain of APP23 mice by adeno-associated viral therapy significantly reduced A β pathology and gliosis and improved cognitive functions. These effects correlated with increased levels of 24-OHC in the brain regions under study, but no changes in total cholesterol levels were observed in these mice [163], suggesting that the effects may be unrelated to CYP46-mediated cholesterol removal from cells. In fact, oxysterols are not only intermediates in the cholesterol elimination pathway but also constitute important signaling molecules. 24-OHC serves as an activator of nuclear transcription factors, liver X receptors α and β [7,164], which increase the expression of cholesterol transport genes [165,166] including ABCA1 in both neurons and glia [167] and ApoE in astrocytes [31,168]. ABCA1 mediates cellular cholesterol efflux in the brain and influences whole-brain cholesterol homeostasis. It has been shown *in vitro* that ABCA1 actively eliminates 24-OHC in the presence of HDL as a lipid acceptor and protects neuronal cells from the toxic effects of 24-OHC accumulation [169]. Previous studies showed that the exposure of SH-SY5Y human neuroblastoma cells to physiological concentrations of 24-OHC led to a 90% loss in cell viability [170]. This last result is in contrast to the above-mentioned studies showing improved cognition and reduced amyloid load in mice with increased Cyp46A1 levels, leaving open the possibility that 24-OHC may produce different, even opposite, effects depending on levels and cell types.

Specific inactivation of ABCA1 in the mouse brain changes synaptic transmission and sensorimotor behavior [171]. Recent experiments *in vivo* showed that mice lacking brain ABCA1 exhibit cortical astrogliosis, increased inflammatory gene expression as well as activation of mitogen-activated protein kinases (MAPKs) following acute lipopolysaccharide (LPS) administration. Mice lacking neuronal ABCA1 develop astrogliosis but show no change in inflammatory gene expression. These findings suggest that coordinated ABCA1 activity across neurons and glial cells influences neuroinflammation and neurodegeneration [172]. Accordingly, other experiments have shown that genetic elimination of either LXR α or β results in age-dependent neurodegenerative changes with accumulation of lipids in neurons, astrocytes, and the meninges [173].

Oxysterols may also modify A β peptide clearance by acting on the blood–brain barrier. 24-OHC and 27-hydroxycholesterol increased the expression of the ABCB1 transporter in brain capillary endothelial cells leading to enhanced A β clearance [174]. Moreover, oxysterols decrease A β peptide generation by brain capillary endothelial cells, by modulating the expression level of APP proteolytic enzymes [175] and possibly by changing the membrane cholesterol content of these cells [176]. Notably, 27-hydroxycholesterol is produced enzymatically in cells outside the nervous system, but it

can enter the brain via the blood–brain barrier (Fig 2) [177], where it may signal the level of plasma cholesterol levels to cells in the brain and impact on the renin–angiotensin system [178]. This system in addition to its roles in salt and water homeostasis and the regulation of blood pressure regulates multiple brain functions such as learning and memory, processing of sensory information, and regulation of emotional responses [179].

Mechanisms by which cholesterol dyshomeostasis affects synaptic activity

Synaptic transmission appears to be particularly sensitive to a disturbance of cholesterol levels, probably because synaptic vesicle release at the presynaptic terminal and the response to signals through neurotransmitter receptors on the postsynaptic side rely entirely on membranous compartments and membrane-bound signaling pathways [180]. Some key aspects of cholesterol function in pre- and postsynaptic activities are schematized in Figure 4.

Role of cholesterol in presynaptic vesicle fusion

For membrane fusion to occur, two bilayers must merge, resulting in extreme structural changes. Membrane curvature is a key determinant for fusion [181] and strongly depends on lipid composition and topology. The merging process between membrane bilayers requires a highly negative curvature [182], and thus, addition of lipids with negative intrinsic curvature facilitates the fusion of bilayers [183].

Cholesterol is a prominent component of synaptic vesicles. It supports intrinsic negative curvature of membranes [184,185] and facilitates the formation of high curvature intermediates during the fusion process. Depletion of cholesterol results in a dose-dependent inhibition of the rate and kinetics of fusion [186]. In addition, cholesterol may favor membrane fusion through its interaction with synaptophysin, an integral membrane protein enriched in synaptic vesicles [187]. Consistent with these data, cholesterol depletion impairs synaptic vesicle exocytosis in cultured neurons [188], greatly reduces Ca²⁺-evoked neurotransmitter release from synaptosomes [189], and alters presynaptic plasticity events [190]. Accordingly, addition of cholesterol to glia- and serum-free neuronal cultures enhances presynaptic transmitter release [191,192].

Furthermore, cholesterol may facilitate fusion by concentrating SNAREs, a highly conserved family of integral membrane proteins involved in synaptic vesicle fusion, at fusion-competent sites [189,193,194]. Cholesterol depletion also affects the ability of synapses to undergo sustained synaptic transmission [192] by compromising the recycling of SV proteins [195]. A thorough understanding of the presynaptic role of cholesterol probably requires more refined methods to manipulate its subcellular levels.

Influence of cholesterol on postsynaptic function

Changes in the number and composition of postsynaptic glutamate receptors contribute to the induction and consolidation of memory formation [196–199]. These changes occur through endocytosis and exocytosis but also through the rapid lateral diffusion of these receptors between synaptic and extrasynaptic areas in the plasma membrane [200,201]. These processes are controlled by receptor interactions with the underlying protein scaffold [202] and by the

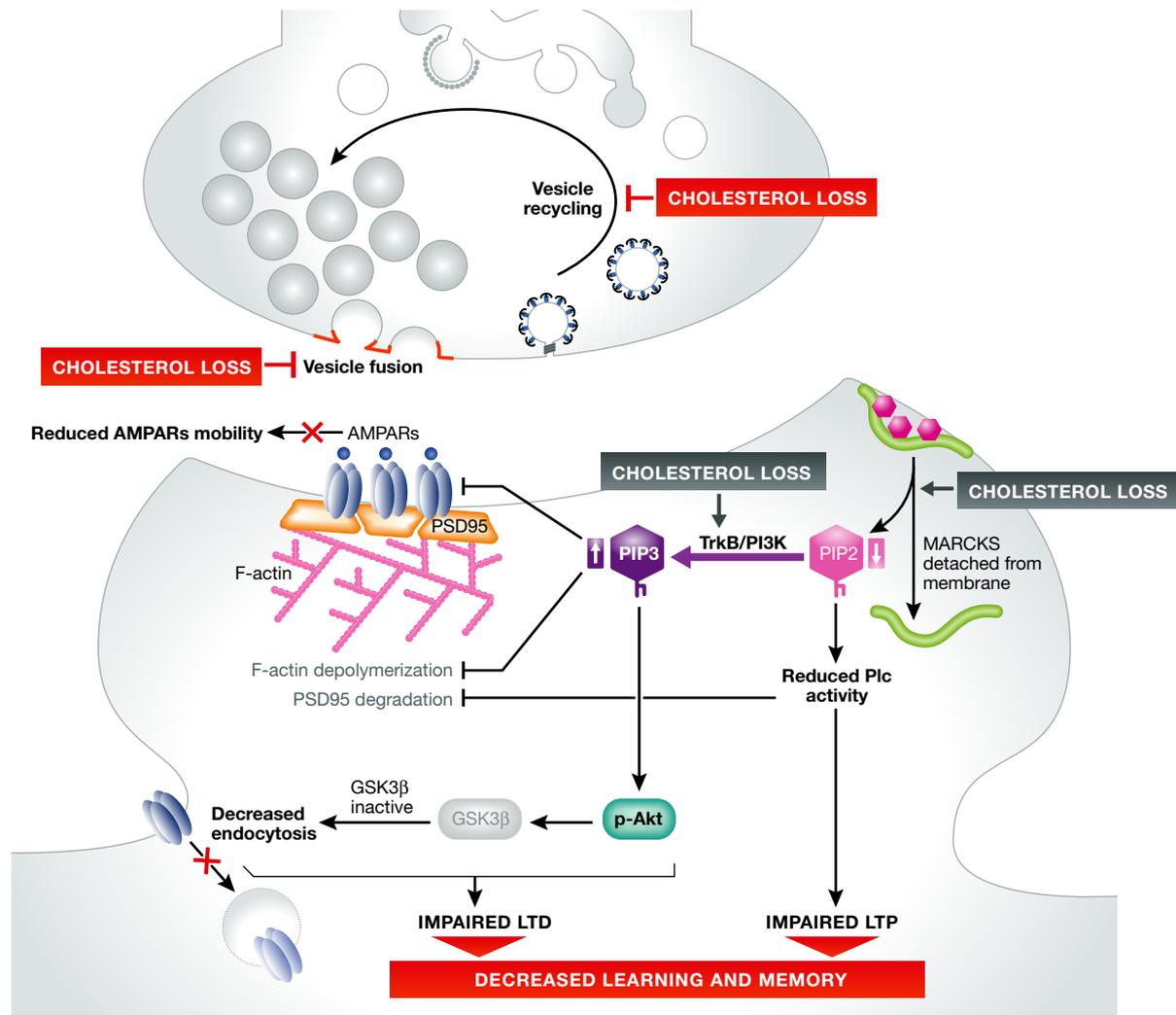


Figure 4. Proposed mechanism to explain how reduced brain cholesterol could underlie poor cognition.

In the presynaptic compartment, cholesterol depletion impairs synaptic vesicle exocytosis probably due to altered membrane curvature and impaired SNARE clusterization at fusion-competent sites. Cholesterol depletion also affects the ability of synapses to undergo sustained synaptic transmission by compromising the recycling of proteins from synaptic vesicles. In the postsynaptic compartment, cholesterol loss leads to MARCKS detachment from the membrane and PIP2 release. Most of the released PIP2 is transformed to PIP3 due to the enhanced TrkB PI3K activity also promoted by cholesterol loss. PIP3 accumulation stabilizes F-actin, blocks AMPARs at the dendritic spines, and also leads to high levels of active p-Akt, which in turn inactivates GSK3β required to promote AMPARs endocytosis. As a consequence of reduced PIP2, low PLCγ activity is also found in old neurons leading to impaired LTP. PLC activity is also required for processes such as actin depolymerization, AKAP150 removal from spines, and PSD95 degradation after LTD induction. Altogether, impaired LTP and LTD in old neurons result in decreased learning and memory.

lipid composition of the subsynaptic membranes [24,203]. It has been shown that depletion of cholesterol destabilizes surface AMPA receptors clustered within lipid rafts in cultured hippocampal neurons [204] and decreases their mobility [203]. In agreement with these observations, we have recently shown that in neurons with low cholesterol levels, AMPA receptors accumulate at the cell surface due to reduced lateral mobility and impaired endocytosis [205], supporting the idea that cholesterol levels influence synaptic activity.

Similarly, the distribution and function of NMDA receptors depends on the lipid environment. Their presence in lipid rafts may facilitate their oligomerization [24]. Cholesterol depletion prevents NMDA-dependent Ca^{2+} influx in cultured hippocampal pyramidal cells [206] and inhibits NMDA-induced long-term potentiation (LTP)

in the hippocampus [207]. Recent studies suggest 24-OHC as a very potent positive allosteric modulator of NMDARs. At sub-micromolar concentrations, 24-OHC potentiated NMDAR-mediated EPSCs in rat hippocampal neurons *in vitro* and enhanced the ability of sub-threshold stimuli to induce LTP in hippocampal slices. In turn, 24-OHC reversed hippocampal LTP deficits induced by the NMDAR channel blocker ketamine. Synthetic drug-like derivatives of 24-OHC are able to restore behavioral and cognitive deficits in rodents treated with NMDAR channel blockers [208].

Previous studies from our laboratory provide evidence for age-dependent changes in the membrane composition of rodent hippocampal neurons. We observed that the membrane concentration of the PI(4,5)P2-clustering molecule MARCKS declines during aging in mice hippocampal neurons. The reduced level of MARCKS lowers

the concentration of PI(4,5)P₂ and reduces PLC γ activity, with a negative impact on learning and memory [209]. More recently, we have shown that cholesterol depletion can trigger the detachment of MARCKS from neuronal membranes in culture, which suggests that the natural occurrence of cholesterol reduction during aging can contribute to the cognitive deficit phenotype of the elderly through, at least in part, this mechanism [205].

Our studies also suggest a link between cholesterol loss and TrkB receptor activation. First, age-related decrease of cholesterol in membranes of hippocampal neurons is accompanied by the recruitment of TrkB receptors to rafts and their phosphorylation *in vivo* and *in vitro* [137]. Second, mild (25%) reductions in membrane cholesterol of cultured neurons activate TrkB and its downstream effector Akt [137,210,211]. Age-dependent accumulation of active Akt occurred in the hippocampus of old mice [137,205,209], which could be restored by replenishment of cholesterol in hippocampal acute slices and in primary neurons with low cholesterol content [205,211]. Long-term depression (LTD) requires dephosphorylation of p-Akt to trigger endocytosis of AMPA receptors, and accumulation of p-Akt in old cells interferes with this process [205]. Electrophysiological recordings from brain slices of old mice and in anesthetized elderly rats demonstrated that the reduced hippocampal LTD associated with age can be rescued by cholesterol perfusion. Accordingly, cholesterol infusion in the lateral ventricle of old animals improved hippocampal-dependent learning and memory in the water maze test [205].

Therapeutic approaches to cholesterol-related brain diseases

Whether disturbances of cholesterol metabolism cause or contribute to brain disease, cholesterol and cholesterol-dependent pathways are obvious targets for therapeutic interventions.

A prime example for a potential therapeutic approach targeting cholesterol is provided by NPC disease. Several studies revealed that administration of a specific form of cyclodextrin can stop the progress of the disease in mouse models of NPC [212,213]. Although the exact mechanism is unknown, the most parsimonious explanation is that the lipophilic molecule equilibrates the cholesterol pools, by extracting cholesterol from sites where it is highly concentrated (such as the endosomal-lysosomal system) and redistributing it to sites with lower content (such as the plasma membrane). Irrespective of the unclear mode of action, a clinical trial exploring cyclodextrin as potential therapeutic approach to NPC patients is currently under way [214]. Interestingly, a recent study revealed that the same form of cyclodextrin has neuroprotective activity in cellular and mouse models of Alzheimer's disease [215].

A helpful indication for clinical trials is the availability of biomarkers that allow for monitoring treatment efficacy in a longitudinal manner. Again, NPC serves as an exemplary case, where progress has been made. Recent studies have shown that the plasma and CSF of NPC patients contain strongly elevated levels of specific oxysterols, namely 3 β ,5 α ,6 β -cholestanetriol and 7-ketocholesterol that are generated by non-enzymatic oxidation [216]. These changes, which are probably caused by elevated oxidative stress in a variety of cell types, are highly specific and can be used to diagnose the disease and to monitor disease progression [217]. Future research should

reveal whether these oxysterols can serve as biomarkers for other neurodegenerative diseases.

There is evidence that LXR activation by the agonist T0901317 reduces neuropathological changes and improves memory in mouse models of experimental dementia [218]. An essential role of LXRs for A β peptide clearance in the APP/PS1 transgenic mouse model has been proposed: activation of LXRs elevates the brain levels of ApoE and ABCA1, resulting in increased amyloid clearance and markedly improved memory formation [219]. Similarly, LXR activation suppressed amyloid deposition and improved memory function in APP23 mice exposed to high fat diet [220], altogether suggesting that LXR activation can play different roles, at the central and systemic levels. It should be mentioned that astrocytic LXR α activation and subsequent release of ApoE by astrocytes plays a role in cholesterol delivery to neuronal cells and has been shown to be critical for the ability of microglia to remove fibrillar A β in response to treatment with LXR activator T0901317 [221]. Note, however, that some LXR agonists are not very specific and may affect multiple molecular targets.

Statins have been assayed for the treatment of neurodegenerative diseases such as Alzheimer's disease, Parkinson's disease, multiple sclerosis, ischemic stroke, and traumatic brain injury. However, the benefits of statins are controversial: some observations suggest that a positive relationship exists and that statins delay both, the onset and progression of dementia [76,222–224], others found a similar risk for dementia among statin and non-statin users [225] and some revealed cognitive impairment by statin treatment [226]. In January 2014, the United States Federal Drug Administration issued a note of advice on statin risks, recognizing the existence of cognitive impairment, such as memory loss, forgetfulness, insomnia, and confusion in some statin users (<http://www.fda.gov/forconsumers/consumerupdates/ucm293330.htm>). In agreement, a recent study showed that long-term oral treatment of mice with atorvastatin, the most widely prescribed statin, leads to significant alterations in behavior and cognition [227]. It is now thought that lipophilicity of the statins may determine the degree of side effects, especially those affecting muscle and central nervous system function, and could also explain the different results obtained in the different clinical trials [228]. It is in fact easy to envision that lipophilic statins can directly alter brain cholesterol levels, leading to dysfunction by the direct reduction of cholesterol in the membrane of astrocytes, neurons, and oligodendrocytes. In addition, one should not forget that statins may also exert beneficial effects through inhibiting the Cox-2 pathway and thus inflammation [229], by affecting the endothelial nitric oxide synthase [230] or by their anti-oxidant properties [231].

Conclusion

In summary, it is clear that a direct disturbance of cholesterol metabolism, for example, by defects in cholesterol synthesizing enzymes or transporters, impairs brain development and function. In addition, changes in cholesterol metabolism in the adult and during aging, and in several age-related neurodegenerative diseases, can directly impact on brain function. Nevertheless, a true causal link between brain cholesterol alterations and late-onset brain dysfunction and brain disease is unproven [232]. In fact, it even remains uncertain to which extent inheritable polymorphisms or mutations in cholesterol pathway genes predispose

Sidebar A: In need of answers

- (i) For which of the major pathologies of the adult brain cholesterol dyshomeostasis is a primary disease-triggering event rather than simply a secondary effect of the disease, such as the multiple defects in cellular homeostasis that occur in the course of a disease (as for example mitochondrial dysfunction, calcium dyshomeostasis, transcriptional dysregulation)? It remains unclear whether polymorphisms in cholesterol synthesis and transport regulatory genes predispose to diseases of late onset like Alzheimer's due to a direct brain cholesterol defect or as a consequence of the accompanying vascular/systemic alterations (or the combination of both).
- (ii) Can the levels of cholesterol-related biomarkers in blood or CSF of patients help to monitor patient-specific changes over time and to define disease progression?
- (iii) Future transcriptional, protein, and lipid analyses should determine the cell types, such as neurons, astrocytes, and oligodendrocytes, in which changes in cholesterol metabolism occur.
- (iv) Should lipophilic, blood–brain barrier permeable, statins be used for the prevention or treatment of brain disorders? This is an important consideration for future clinical trials, in light of the numerous evidences for decreased cholesterol content and metabolism in the brains of the aged, and the reports (and formal warnings from the FDA) of cognitive and mood disorders in statin users.
- (v) Future studies should determine by which of its multiple sites of action statins produce 'beneficial' effects: through reduction of cellular cholesterol/cholesterol metabolites, anti-inflammatory, anti-oxidative stress, or the nitric oxide pathway?

to brain pathology of the adult by a direct perturbation of brain cholesterol homeostasis rather than body cholesterol/metabolic dyshomeostasis indirectly affecting brain function. Like in AD, the changes in brain cholesterol in inherited conditions with symptoms in the adult could be an accompanying process, likely relevant in the stabilization or progression of disease signs, rather than being the cause of the disease. Further progress in the field requires careful cell-specific analyses of cholesterol homeostasis in neurons and glial cells under different pathologic conditions with new tools for cell-specific measurements and manipulations of cholesterol levels *in vivo*.

Conflict of interest

The authors declare that they have no conflict of interest.

References

1. Mañes S, Martínez-A C (2004) Cholesterol domains regulate the actin cytoskeleton at the leading edge of moving cells. *Trends Cell Biol* 14: 275–278
2. Dart C (2010) Lipid microdomains and the regulation of ion channel function. *J Physiol* 588: 3169–3178
3. Lippincott-Schwartz J, Phair RD (2010) Lipids and cholesterol as regulators of traffic in the endomembrane system. *Annu Rev Biophys* 39: 559–578
4. Simons K, Gerl MJ (2010) Revitalizing membrane rafts: new tools and insights. *Nat Rev Mol Cell Biol* 10: 688–699
5. Levitan I, Singh DK, Rosenhouse-Dantsker A (2014) Cholesterol binding to ion channels. *Front Physiol* 5: 65
6. McLean KJ, Hans M, Munro AW (2012) Cholesterol, an essential molecule: diverse roles involving cytochrome P450 enzymes. *Biochem Soc Trans* 40: 587–593
7. Janowski BA, Grogan MJ, Jones SA, Wisely GB, Kliewer SA, Corey EJ, Mangelsdorf DJ (1999) Structural requirements of ligands for the oxysterol liver X receptors LXRA and LXRbeta. *Proc Natl Acad Sci USA* 96: 266–271
8. Björkhem I (2006) Crossing the barrier: oxysterols as cholesterol transporters and metabolic modulators in the brain. *J Intern Med* 260: 493–508
9. Radhakrishnan A, Ikeda Y, Kwon HJ, Brown MS, Goldstein JL (2007) Sterol-regulated transport of SREBPs from endoplasmic reticulum to Golgi: oxysterols block transport by binding to Insig. *Proc Natl Acad Sci USA* 104: 6511–6518
10. Gill S, Chow R, Brown AJ (2008) Sterol regulators of cholesterol homeostasis and beyond: the oxysterol hypothesis revisited and revised. *Prog Lipid Res* 47: 391–404
11. Miyoshi N, Iuliano L, Tomono S, Ohshima H (2014) Implications of cholesterol autoxidation products in the pathogenesis of inflammatory diseases. *Biochem Biophys Res Commun* 446: 702–708
12. Berg JM (2002) *The Complex Regulation of Cholesterol Biosynthesis Takes Place at Several Levels*. Biochemistry, 5th edn. New York: W.H. Freeman
13. Lev S (2010) Non-vesicular lipid transport by lipid-transfer proteins and beyond. *Nat Rev Mol Cell Biol* 10: 739–750
14. Ikonen E (2008) Cellular cholesterol trafficking and compartmentalization. *Nat Rev Mol Cell Biol* 9: 125–138
15. Storch J, Xu Z (2009) Niemann-Pick C2 (NPC2) and intracellular cholesterol trafficking. *Biochim Biophys Acta* 1791: 671–678
16. Brown MS, Goldstein JL (1999) A proteolytic pathway that controls the cholesterol content of membranes, cells, and blood. *Proc Natl Acad Sci USA* 96: 11041–11048
17. Mondal M, Mesmin B, Mukherjee S, Maxfield FR (2009) Sterols are mainly in the cytoplasmic leaflet of the plasma membrane and the endocytic recycling compartment in CHO cells. *Mol Biol Cell* 20: 581–588
18. Lingwood D, Simons K (2010) Lipid rafts as a membrane-organizing principle. *Science* 327: 46–50
19. van Meer G (2011) Dynamic transbilayer lipid asymmetry. *Cold Spring Harb Perspect Biol* 3: a004671
20. Gil C, Cubí R, Blasi J, Aguilera J (2006) Synaptic proteins associate with a sub-set of lipid rafts when isolated from nerve endings at physiological temperature. *Biochem Biophys Res Commun* 348: 1334–1342
21. Wasser CR, Kavalali ET (2009) Leaky synapses: regulation of spontaneous neurotransmission in central synapses. *Neuroscience* 158: 177–188
22. Mailman T, Hariharan M, Karten B (2011) Inhibition of neuronal cholesterol biosynthesis with lovastatin leads to impaired synaptic vesicle release even in the presence of lipoproteins or geranylgeraniol. *J Neurochem* 119: 1002–1015
23. Suzuki T (2002) Lipid rafts at postsynaptic sites: distribution, function and linkage to postsynaptic density. *Neurosci Res* 44: 1–9
24. Allen JA, Halverson-Tamboli RA, Rasenick MM (2007) Lipid raft microdomains and neurotransmitter signalling. *Nat Rev Neurosci* 8: 128–140
25. Dietschy JM, Turley SD (2004) Thematic review series: brain Lipids. Cholesterol metabolism in the central nervous system during early development and in the mature animal. *Lipid Res* 45: 1375–1397

26. Pfrieger FW (2003) Cholesterol homeostasis and function in neurons of the central nervous system. *Cell Mol Life Sci* 60: 1158–1171
27. Björkhem I, Meaney S (2004) Brain cholesterol: long secret life behind a barrier. *Arterioscler Thromb Vasc Biol* 24: 806–815
28. Russell DW, Halford RW, Ramirez DM, Shah R, Kotti T (2009) Cholesterol 24-hydroxylase: an enzyme of cholesterol turnover in the brain. *Annu Rev Biochem* 78: 1017–1040
29. Quan G, Xie C, Dietschy JM, Turley SD (2003) Ontogenesis and regulation of cholesterol metabolism in the central nervous system of the mouse. *Brain Res Dev Brain Res* 146: 87–98
30. Thelen KM, Falkai P, Bayer TA, Lütjohann D (2006) Cholesterol synthesis rate in human hippocampus declines with aging. *Neurosci Lett* 403: 15–19
31. Pfrieger FW, Ungerer N (2011) Cholesterol metabolism in neurons and astrocytes. *Prog Lipid Res* 50: 357–371
32. Porter FD, Herman GE (2011) Malformation syndromes caused by disorders of cholesterol synthesis. *J Lipid Res* 52: 6–34
33. Kanungo S, Soares N, He M, Steiner RD (2013) Sterol metabolism disorders and neurodevelopment—an update. *Dev Disabil Res Rev* 17: 197–210
34. Vanier MT (2013) Niemann-Pick diseases. *Handb Clin Neurol* 113: 1717–1721
35. Waterham HR, Hennekam RC (2012) Mutational spectrum of Smith-Lemli-Opitz syndrome. *Am J Med Genet C Semin Med Genet* 160: 263–284
36. De Barber AE, Eroglu Y, Merkens LS, Pappu AS, Steiner RD (2011) Smith-Lemli-Opitz syndrome. *Expert Rev Mol Med* 13: e24
37. Diaz-Stransky A, Tierney E (2012) Cognitive and behavioral aspects of Smith-Lemli-Opitz syndrome. *Am J Med Genet C Semin Med Genet* 160: 295–300
38. Nowaczyk MJ, Irons MB (2012) Smith-Lemli-Opitz syndrome: phenotype, natural history, and epidemiology. *Am J Med Genet C Semin Med Genet* 160: 250–262
39. Cunniff C, Kratz LE, Moser A, Natowicz MR, Kelley RI (1997) Clinical and biochemical spectrum of patients with RSH/Smith-Lemli-Opitz syndrome and abnormal cholesterol metabolism. *Am J Med Genet* 68: 263–269
40. Cooper MK, Porter JA, Young KE, Beachy PA (1998) Teratogen-mediated inhibition of target tissue response to Shh signaling. *Science* 280: 1603–1607
41. Koide T, Hayata T, Cho K W Y (2006) Negative regulation of Hedgehog signaling by the cholesterologenic enzyme 7-dehydrocholesterol reductase. *Development* 133: 2395–2405
42. Porter JA, Young KE, Beachy PA (1996) Cholesterol modification of hedgehog signaling proteins in animal development. *Science* 274: 255–259
43. Gallet A, Ruel L, Staccini-Lavenant L, Théron PP (2005) Cholesterol modification is necessary for controlled planar long-range activity of Hedgehog in *Drosophila* epithelia. *Development* 133: 407–418
44. Zerenturk EJ, Sharpe LJ, Ikonen E, Brown AJ (2013) Desmosterol and DHCR24: unexpected new directions for a terminal step in cholesterol synthesis. *Prog Lipid Res* 52: 666–680
45. Klahre U, Noguchi T, Fujioka S, Takatsuto S, Yokota T (1998) The Arabidopsis DIMINUTO/DWARF1 gene encodes a protein involved in steroid synthesis. *Plant Cell* 10: 1677–1690
46. Schaller H (2003) The role of sterols in plant growth and development. *Prog Lipid Res* 42: 163–175
47. Wu C, Miloslavskaya I, Demontis S, Maestro R, Galaktionov K (2004) Regulation of cellular response to oncogenic and oxidative stress by Seladin-1. *Nature* 432: 640–645
48. Greeve I, Hermans-Borgmeyer C, Brellinger D, Kasper T, Gomez-Isla T, Behl C, Levkau B, Nitsch RM (2000) The human DIMINUTO/DWARF1 homolog seladin-1 confers resistance to Alzheimer's disease-associated neurodegeneration and oxidative stress. *J Neurosci* 20: 7345–7352
49. Cramer A, Biondi E, Kuehne K, Lütjohann D, Thelen KM, Perga S, Dotti CG, Nitsch RM, Ledesma MD, Mohajeri MH (2006) The role of seladin-1/DHCR24 in cholesterol biosynthesis, APP processing and Abeta generation in vivo. *EMBO J* 25: 432–443
50. Vanier MT (2010) Niemann-Pick disease type C. *Orphanet J Rare Dis* 3: 16
51. Patterson MC, Hendriksz CJ, Walterfang M, Sedel F, Vanier MT, Wijburg F (2012) Recommendations for the diagnosis and management of Niemann-Pick disease type C: an update. *Mol Genet Metab* 106: 330–344
52. Walkley SU, Suzuki K (2004) Consequences of NPC1 and NPC2 loss of function in mammalian neurons. *Biochim Biophys Acta* 1685: 48–62
53. Kwon HJ, Abi-Mosleh L, Wang ML, Deisenhofer J, Goldstein JL, Brown MS, Infante RE (2009) Structure of N-terminal domain of NPC1 reveals distinct subdomains for binding and transfer of cholesterol. *Cell* 137: 1213–1224
54. Liscum L (2000) Niemann-Pick type C mutations cause lipid traffic jam. *Traffic* 1: 218–225
55. Vance JE, Karten B (2014) Niemann-Pick C disease and mobilization of lysosomal cholesterol by cyclodextrin. *J Lipid Res* 55: 1609–1621
56. Pentchev PG, Comly ME, Kruth HS, Vanier MT, Wenger DA, Patel S, Brady RO (1985) A defect in cholesterol esterification in Niemann-Pick disease (type C) patients. *Proc Natl Acad Sci USA* 8: 8247–8251
57. Lloyd-Evans E, Morgan AJ, He X, Smith DA, Elliot-Smith E, Sillence DJ, Churchill GC, Schuchman EH, Galione A, Platt FM (2008) Niemann-Pick disease type C1 is a sphingosine storage disease that causes deregulation of lysosomal calcium. *Nat Med* 14: 1247–1255
58. Lloyd-Evans E, Platt FM (2010) Lipids on trial: the search for the offending metabolite in Niemann-Pick type C disease. *Traffic* 8: 419–428
59. Liscum L, Ruggiero RM, Faust JR (1989) The intracellular transport of low density lipoprotein-derived cholesterol is defective in Niemann-Pick type C fibroblasts. *J Cell Biol* 108: 1625–1636
60. Karten B, Vance DE, Campenot RB, Vance JE (2002) Cholesterol accumulates in cell bodies, but is decreased in distal axons, of Niemann-Pick C1-deficient neurons. *J Neurochem* 83: 1154–1163
61. Ko DC, Gordon MD, Jin JY, Scott MP (2001) Dynamic movements of organelles containing Niemann-Pick C1 protein: NPC1 involvement in late endocytic events. *Mol Biol Cell* 12: 601–614
62. Karten B, Campenot RB, Vance DE, Vance JE (2006) The Niemann-Pick C1 protein in recycling endosomes of presynaptic nerve terminals. *J Lipid Res* 47: 504–514
63. Vance JE, Hayashi H, Karten B (2005) Cholesterol homeostasis in neurons and glial cells. *Semin Cell Dev Biol* 16: 193–212
64. Chahrouh M, Jung SY, Shaw C, Zhou X, Wong ST, Qin J, Zoghbi HY (2008) MeCP2, a key contributor to neurological disease, activates and represses transcription. *Science* 320: 1224–1229
65. Buchovecky CM, Turley SD, Brown HM, Kyle SM, McDonald JG, Liu B, Pieper AA, Huang W, Katz DM, Russell DW et al (2013) A suppressor screen in Mecp2 mutant mice implicates cholesterol metabolism in Rett syndrome. *Nat Genet* 45: 1013–1020
66. Joshi HN, Fakes MG, Serajuddin ATM (1999) Differentiation of 3-hydroxy-3-methylglutaryl-coenzyme A reductase inhibitors by their relative lipophilicity. *Pharm Pharmacol Commun* 5: 269–271

67. Li Y, Wang H, Muffat J, Cheng AW, Orlando DA, Lovén J, Kwok SM, Feldman DA, Bateup HS, Gao Q *et al* (2013) Global transcriptional and translational repression in human-embryonic-stem-cell-derived Rett syndrome neurons. *Cell Stem Cell* 13: 446–458
68. Saunders AM, Strittmatter WJ, Schmechel D, George-Hyslop PH, Pericak-Vance MA, Joo SH, Rosi BL, Gusella JF, Crapper-MacLachlan DR, Alberts MJ (1993) Association of apolipoprotein E allele epsilon 4 with late-onset familial and sporadic Alzheimer's disease. *Neurology* 43: 1467–1472
69. Schmechel DE, Saunders AM, Strittmatter WJ, Crain BJ, Hulette CM, Joo SH, Pericak-Vance MA, Goldgaber D, Roses AD (1993) Increased amyloid beta-peptide deposition in cerebral cortex as a consequence of apolipoprotein E genotype in late-onset Alzheimer disease. *Proc Natl Acad Sci USA* 90: 9649–9653
70. Ordovas JM, Litwack-Klein L, Wilson PW, Schaefer MM, Schaefer EJ (1987) Apolipoprotein E isoform phenotyping methodology and population frequency with identification of apoE1 and apoE5 isoforms. *J Lipid Res* 28: 371–380
71. Lehtinen S, Lehtimäki T, Sisto T, Salenius JP, Nikkilä M, Jokela H, Koivula T, Ebeling F, Ehnholm C (1995) Apolipoprotein E polymorphism, serum lipids, myocardial infarction and severity of angiographically verified coronary artery disease in men and women. *Atherosclerosis* 114: 83–91
72. Howard BV, Gidding SS, Liu K (1998) Association of apolipoprotein E phenotype with plasma lipoproteins in African-American and white young adults. The CARDIA Study. Coronary Artery Risk Development in Young Adults. *Am J Epidemiol* 148: 859–868
73. Sparks DL, Scheff SW, Hunsaker JC 3rd, Liu H, Landers T, Gross DR (1994) Induction of Alzheimer-like beta-amyloid immunoreactivity in the brains of rabbits with dietary cholesterol. *Exp Neurol* 126: 88–94
74. Zambón D, Quintana M, Mata P, Alonso R, Benavent J, Cruz-Sánchez F, Gich J, Pocióvi M, Civeira F, Capurro S *et al* (2010) Higher incidence of mild cognitive impairment in familial hypercholesterolemia. *Am J Med* 123: 267–274
75. Wolozin B, Kellman W, Rousseau P, Celesia GG, Siegel G (2000) Decreased prevalence of Alzheimer disease associated with 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitors. *Arch Neurol* 57: 1439–1443
76. Jick H, Zornberg GL, Jick SS, Seshadri S, Drachman DA (2000) Statins and the risk of dementia. *Lancet* 356: 1627–1631
77. Reed B, Villeneuve S, Mack W, DeCarli C, Chui HC, Jagust W (2014) Associations between serum cholesterol levels and cerebral amyloidosis. *JAMA Neurol* 71: 195–200
78. Altman R, Rutledge JC (2010) The vascular contribution to Alzheimer's disease. *Clin Sci (Lond)* 119: 407–421
79. Tenenholz Grinberg L, Thal DR (2010) Vascular pathology in the aged human brain. *Acta Neuropathol* 119: 277–290
80. Vasilevko V, Passos GF, Quiring D, Head E, Kim RC, Fisher M, Cribbs DH (2010) Aging and cerebrovascular dysfunction: contribution of hypertension, cerebral amyloid angiopathy, and immunotherapy. *Ann N Y Acad Sci* 207: 58–70
81. de la Torre JC (2004) Is Alzheimer's disease a neurodegenerative or a vascular disorder? Data, dogma, and dialectics. *Lancet Neurol* 3: 184–190
82. de la Torre JC (2006) How do heart disease and stroke become risk factors for Alzheimer's disease? *Neurol Res* 28: 637–644
83. Newman AB, Fitzpatrick AL, Lopez O, Jackson S, Lyketsos C, Jagust W, Ives D, Dekosky ST, Kuller LH (2005) Dementia and Alzheimer's disease incidence in relationship to cardiovascular disease in the Cardiovascular Health Study cohort. *J Am Geriatr Soc* 53: 1101–1107
84. Honig LS, Kukull W, Mayeux R (2005) Atherosclerosis and AD: analysis of data from the US National Alzheimer's Coordinating Center. *Neurology* 64: 494–500
85. Bell RD (2012) The imbalance of vascular molecules in Alzheimer's disease. *J Alzheimers Dis* 32: 699–709
86. Orsucci D, Mancuso M, Ienco EC, Simoncini C, Siciliano G, Bonuccelli U (2013) Vascular factors and mitochondrial dysfunction: a central role in the pathogenesis of Alzheimer's disease. *Curr Neurol Res* 10: 76–80
87. Zhiyou C, Yong Y, Shanquan S, Jun Z, Liangguo H, Ling Y, Jieying L (2009) Upregulation of BACE1 and beta-amyloid protein mediated by chronic cerebral hypoperfusion contributes to cognitive impairment and pathogenesis of Alzheimer's disease. *Neurochem Res* 34: 1226–1235
88. Kitaguchi H, Tomimoto H, Ihara M, Shibata M, Uemura K, Kalaria RN, Kihara T, Asada-Utsugi M, Kinoshita A, Takahashi R (2009) Chronic cerebral hypoperfusion accelerates amyloid beta deposition in APPS-wind transgenic mice. *Brain Res* 1294: 202–210
89. Poli G, Sottero B, Gargiulo S, Leonarduzzi G (2009) Cholesterol oxidation products in the vascular remodeling due to atherosclerosis. *Mol Aspects Med* 30: 180–189
90. Janson J, Laedtke T, Parisi JE, O'Brien P, Petersen RC, Butler PC (2004) Increased risk of type 2 diabetes in Alzheimer disease. *Diabetes* 53: 474–481
91. Kruit JK, Brunham LR, Verchere CB, Hayden MR (2010) HDL and LDL cholesterol significantly influence beta-cell function in type 2 diabetes mellitus. *Curr Opin Lipidol* 21: 178–185
92. Ledesma MD, Dotti CG (2012) Peripheral cholesterol, metabolic disorders and Alzheimer's disease. *Front Biosci (Elite Ed)* 4: 181–194
93. Bu G (2009) Apolipoprotein E and its receptors in Alzheimer's disease: pathways, pathogenesis and therapy. *Nat Rev Neurosci* 10: 333–344
94. Gong JS, Kobayashi M, Hayashi H, Zou K, Sawamura N, Fujita SC, Yanagisawa K, Michikawa M (2002) Apolipoprotein E (ApoE) isoform-dependent lipid release from astrocytes prepared from human ApoE3 and ApoE4 knock-in mice. *J Biol Chem* 277: 29919–29926
95. Hayashi H, Campenot RB, Vance DE, Vance JE (2007) Apolipoprotein E-containing lipoproteins protect neurons from apoptosis via a signaling pathway involving low-density lipoprotein receptor-related protein-1. *J Neurosci* 27: 1933–1941
96. Martins LJ, Hone E, Foster JK, Sünram-Lea SI, Gnjec A, Fuller SJ, Nolan D, Gandy SE, Martins RN (2006) Apolipoprotein E, cholesterol metabolism, diabetes, and the convergence of risk factors for Alzheimer's disease and cardiovascular disease. *Mol Psychiatry* 11: 721–736
97. Castellano JM, Kim J, Stewart FR, Jiang H, DeMattos RB, Patterson BW, Fagan AM, Morris JC, Mawuenyega KG, Cruchaga C *et al* (2011) Human apoE isoforms differentially regulate brain amyloid- β peptide clearance. *Sci Transl Med* 3: 89ra57
98. Cramer PE, Cirrito JR, Wesson DW, Lee CY, Karlo JC, Zinn AE, Casali BT, Restivo JL, Goebel WD, James MJ *et al* (2012) ApoE-directed therapeutics rapidly clear β -amyloid and reverse deficits in AD mouse models. *Science* 335: 1503–1506
99. Bien-Ly N, Gillespie AK, Walker D, Yoon SY, Huang Y (2012) Reducing human apolipoprotein E levels attenuates age-dependent A β accumulation in mutant human amyloid precursor protein transgenic mice. *J Neurosci* 32: 4803–4811
100. Kanekiyo T, Xu H, Bu G (2014) ApoE and A β in Alzheimer's disease: accidental encounters or partners? *Neuron* 81: 740–754

101. Zlokovic BV (2013) Cerebrovascular effects of apolipoprotein E: implications for Alzheimer disease. *JAMA Neurol* 70: 440–444
102. Lambert JC, Amouyel P (2011) Genetics of Alzheimer's disease: new evidences for an old hypothesis? *Curr Opin Genet Dev* 21: 295–301
103. Silkenen JR, Schwachau GB, Rosenberg ME (1994) The role of clusterin in tissue injury. *Biochem Cell Biol* 72: 483–488
104. Humphreys DT, Carver JA, Easterbrook-Smith SB, Wilson MR (1999) Clusterin has chaperone-like activity similar to that of small heat shock proteins. *J Biol Chem* 274: 6875–6881
105. Miwa Y, Takiuchi S, Kamide K, Yoshii M, Horio T, Tanaka C, Banno M, Miyata T, Sasaguri T, Kawano Y (2005) Insertion/deletion polymorphism in clusterin gene influences serum lipid levels and carotid intima-media thickness in hypertensive Japanese females. *Biochem Biophys Res Commun* 331: 1587–1593
106. Rosenberg ME, Girton R, Finkel D, Chmielewski D, Barrie A 3rd, Witte DP, Zhu G, Bissler JJ, Harmony JA, Aronow BJ (2002) Apolipoprotein J/clusterin prevents a progressive glomerulopathy of aging. *Mol Cell Biol* 22: 1893–1902
107. Lee JY, Parks JS (2005) ATP-binding cassette transporter A1 and its role in HDL formation. *Curr Opin Lipidol* 16: 19–25
108. Grimm MO, Rothhaar TL, Hartmann T (2012) The role of APP proteolytic processing in lipid metabolism. *Exp Brain Res* 217: 365–375
109. Cutler RG, Kelly J, Storie K, Pedersen WA, Tammara A, Hatanpaa K, Troncoso JC, Mattson MP (2004) Involvement of oxidative stress-induced abnormalities in ceramide and cholesterol metabolism in brain aging and Alzheimer's disease. *Proc Natl Acad Sci USA* 101: 2070–2075
110. Mulder M, Ravid R, Swaab DF, de Kloet ER, Haasdijk ED, Julk J, van der Boom JJ, Havekes LM (1998) Reduced levels of cholesterol, phospholipids, and fatty acids in cerebrospinal fluid of Alzheimer disease patients are not related to apolipoprotein E4. *Alzheimer Dis Assoc Disord* 12: 198–203
111. Kölsch H, Heun R, Jessen F, Popp J, Hentschel F, Maier W, Lütjohann D (2010) Alterations of cholesterol precursor levels in Alzheimer's disease. *Biochim Biophys Acta* 1801: 945–950
112. Söderberg M, Edlund C, Kristensson K, Dallner G (1990) Lipid compositions of different regions of the human brain during aging. *J Neurochem* 54: 415–423
113. Mason RP, Shoemaker WJ, Shajenko L, Chambers TE, Herbette LG (1992) Evidence for changes in the Alzheimer's disease brain cortical membrane structure mediated by cholesterol. *Neurobiol Aging* 13: 413–419
114. Roher AE, Weiss N, Kokjohn TA, Kuo YM, Kalback W, Anthony J, Watson D, Luehrs DC, Sue L, Walker D et al (2002) Increased A beta peptides and reduced cholesterol and myelin proteins characterize white matter degeneration in Alzheimer's disease. *Biochemistry* 41: 11080–11090
115. Papassotiropoulos A, Lütjohann D, Bagli M, Locatelli S, Jessen F, Buschfort R, Ptok U, Björkhem I, von Bergmann K, Heun R (2002) 24S-hydroxycholesterol in cerebrospinal fluid is elevated in early stages of dementia. *J Psychiatr Res* 36: 27–32
116. Molander-Melin M, Blennow K, Bogdanovic N, Dellheden B, Månsson JE, Fredman P (2005) Structural membrane alterations in Alzheimer brains found to be associated with regional disease development; increased density of gangliosides GM1 and GM2 and loss of cholesterol in detergent-resistant membrane domains. *J Neurochem* 92: 171–182
117. Wang W, Mutka AL, Zmrzljak UP, Rozman D, Tanila H, Gylling H, Remes AM, Huttunen HJ, Ikonen E (2014) Amyloid precursor protein α - and β -cleaved ectodomains exert opposing control of cholesterol homeostasis via SREBP2. *FASEB J* 28: 849–860
118. Gylys KH, Fein JA, Yang F, Miller CA, Cole GM (2007) Increased cholesterol in Abeta-positive nerve terminals from Alzheimer's disease cortex. *Neurobiol Aging* 28: 8–17
119. Mori T, Paris D, Town T, Rojiani AM, Sparks DL, Delledonne A, Crawford F, Abdullah LI, Humphrey JA, Dickson DW et al (2001) Cholesterol accumulates in senile plaques of Alzheimer disease patients and in transgenic APP(SW) mice. *J Neuropathol Exp Neurol* 60: 778–785
120. Pierrot N, Tyteca D, D'auria L, Dewachter I, Gaillly P, Hendrickx A, Tasi-aux B, Haylani LE, Muls N, N'kuli F et al (2013) Amyloid precursor protein controls cholesterol turnover needed for neuronal activity. *EMBO Mol Med* 5: 608–625
121. Beel AJ, Sakakura M, Barrett PJ, Sanders CR (2010) Direct binding of cholesterol to the amyloid precursor protein: an important interaction in lipid-Alzheimer's disease relationships? *Biochim Biophys Acta* 1801: 975–982
122. Lütjohann D, Papassotiropoulos A, Björkhem I, Locatelli S, Bagli M, Oehring RD, Schlegel U, Jessen F, Rao ML, von Bergmann K et al (2000) Plasma 24S-hydroxycholesterol (cerebrosterol) is increased in Alzheimer and vascular demented patients. *J Lipid Res* 41: 195–198
123. Bogdanovic N, Bretillon L, Lund EG, Diczfalusy U, Lannfelt L, Winblad B, Russell DW, Björkhem I (2001) On the turnover of brain cholesterol in patients with Alzheimer's disease. Abnormal induction of the cholesterol-catabolic enzyme CYP46 in glial cells. *Neurosci Lett* 314: 45–48
124. Evangelisti E, Zampagni M, Cascella R, Becatti M, Fiorillo C, Caselli A, Bagnoli S, Nacmias B, Cecchi C (2014) Plasma membrane injury depends on bilayer lipid composition in Alzheimer's disease. *J Alzheimers Dis* 41: 289–300
125. Ohyama Y, Meaney S, Heverin M, Ekström L, Brafman A, Shafir M, Andersson U, Olin M, Eggertsen G, Diczfalusy U et al (2006) Studies on the transcriptional regulation of cholesterol 24-hydroxylase (CYP46A1): marked insensitivity toward different regulatory axes. *J Biol Chem* 281: 3810–3820
126. Bordji K, Becerril-Ortega J, Buisson A (2011) Synapses, NMDA receptor activity and neuronal A β production in Alzheimer's disease. *Rev Neurosci* 22: 285–294
127. Svennerholm L, Bostrom K, Helander CG, Jungbjer B (1991) Membrane lipids in the aging human brain. *J Neurochem* 56: 2051–2059
128. Svennerholm L, Boström K, Jungbjer B, Olsson L (1994) Membrane lipids of adult human brain: lipid composition of frontal and temporal lobe in subjects of age 20 to 100 years. *J Neurochem* 63: 1802–1811
129. Svennerholm L, Bostrom K, Jungbjer B (1997) Changes in weight and compositions of major membrane components of human brain during the span of adult human life of Swedes. *Acta Neuropathol* 94: 345–352
130. Fabelo N, Martín V, Santpere G, Marín R, Torrent L, Ferrer I, Díaz M (2012) Evidence for premature lipid raft aging in APP/PS1 double-transgenic mice, a model of familial Alzheimer disease. *J Neuropathol Exp Neurol* 71: 868–881
131. Sodero AO, Trovò L, Iannilli F, Van Veldhoven P, Dotti CG, Martin MG (2011) Regulation of tyrosine kinase B activity by the Cyp46/cholesterol loss pathway in mature hippocampal neurons: relevance for neuronal survival under stress and in aging. *J Neurochem* 116: 747–755
132. Abad-Rodriguez J, Ledesma MD, Craessaerts K, Perga S, Medina M, Delacourte A, Dingwall C, De Strooper B, Dotti CG (2004) Neuronal membrane cholesterol loss enhances amyloid peptide generation. *J Cell Biol* 167: 953–960
133. Ledesma MD, Abad-Rodriguez J, Galvan C, Biondi E, Navarro P, Delacourte A, Dingwall C, Dotti CG (2003) Raft disorganization leads to

- reduced plasmin activity in Alzheimer's disease brains. *EMBO Rep* 4: 1190–1196
134. Calleros L, Lasa M, Toro MJ, Chiloeches A (2006) Low cell cholesterol levels increase NF κ B activity through a p38 MAPK-dependent mechanism. *Cell Signal* 18: 2292–2301
 135. Zampagni M, Evangelisti E, Cascella R, Liguri G, Becatti M, Pensalfini A, Uberti D, Cenini G, Memo M, Bagnoli S *et al* (2010) Lipid rafts are primary mediators of amyloid oxidative attack on plasma membrane. *J Mol Med (Berl)* 88: 597–608
 136. Pensalfini A, Zampagni M, Liguri G, Becatti M, Evangelisti E, Fiorillo C, Bagnoli S, Cellini E, Nacmias B, Sorbi S *et al* (2011) Membrane cholesterol enrichment prevents A β -induced oxidative stress in Alzheimer's fibroblasts. *Neurobiol Aging* 32: 210–222
 137. Martin MG, Perga S, Trovo L, Rasola A, Holm P, Rantamaki T, Dotti CG (2008) Cholesterol loss enhances TrkB signaling in hippocampal neurons aging in vitro. *Mol Biol Cell* 19: 2101–2112
 138. Sodero AO, Vriens J, Ghosh D, Stegner D, Brachet A, Pallotto M, Sassòè-Pognetto M, Brouwers JF, Helms JB, Nieswandt B *et al* (2012) Cholesterol loss during glutamate-mediated excitotoxicity. *EMBO J* 31: 1764–1773
 139. Teunissen CE, Floris S, Sonke M, Dijkstra CD, De Vries HE, Lütjohann D (2007) 24S-hydroxycholesterol in relation to disease manifestations of acute experimental autoimmune encephalomyelitis. *J Neurosci Res* 85: 1499–1505
 140. Cartagena CM, Ahmed F, Burns MP, Pajoohesh-Ganji A, Pak DT, Faden AI, Rebeck GW (2008) Cortical injury increases cholesterol 24S hydroxylase (Cyp46) levels in the rat brain. *J Neurotrauma* 25: 1087–1098
 141. Valenza M, Rigamonti D, Goffredo D, Zuccato C, Fenu S, Jamot L, Strand A, Tarditi A, Woodman B, Racchi M *et al* (2005) Dysfunction of the cholesterol biosynthetic pathway in Huntington's disease. *J Neurosci* 25: 9932–9939
 142. Leoni V, Caccia C (2014) Study of cholesterol metabolism in Huntington's disease. *Biochem Biophys Res Commun* 446: 697–701
 143. Valenza M, Leoni V, Karasinska JM, Petricca L, Fan J, Carroll J, Pouladi MA, Fossale E, Nguyen HP, Riess O *et al* (2010) Cholesterol defect is marked across multiple rodent models of Huntington's disease and is manifest in astrocytes. *J Neurosci* 30: 10844–10850
 144. Trushina E, Singh RD, Dyer RB, Cao S, Shah VH, Parton RG, Pagano RE, McMurray CT (2006) Mutant huntingtin inhibits clathrin-independent endocytosis and causes accumulation of cholesterol in vitro and in vivo. *Hum Mol Genet* 15: 3578–3591
 145. Fütter M, Diekmann H, Schoenmakers E, Sadiq O, Chatterjee K, Rubinsztein DC (2009) Wild-type but not mutant huntingtin modulates the transcriptional activity of liver X receptors. *J Med Genet* 46: 438–446
 146. Davie CA (2008) A review of Parkinson's disease. *Br Med Bull* 86: 109–127
 147. Bar-On P, Rockenstein E, Adame A, Ho G, Hashimoto M, Masliah E (2006) Effects of the cholesterol-lowering compound methyl-beta-cyclodextrin in models of alpha-synucleinopathy. *J Neurochem* 98: 1032–1045
 148. Koob AO, Ubhi K, Paulsson JF, Kelly J, Rockenstein E, Mante M, Adame A, Masliah E (2010) Lovastatin ameliorates alpha-synuclein accumulation and oxidation in transgenic mouse models of alpha-synucleinopathies. *Exp Neurol* 221: 267–274
 149. Ghosh A, Roy A, Matras J, Brahmachari S, Gendelman HE, Pahan K (2009) Simvastatin inhibits the activation of p21ras and prevents the loss of dopaminergic neurons in a mouse model of Parkinson's disease. *J Neurosci* 29: 13543–13556
 150. Rajanikant GK, Zemke D, Kassab M, Majid A (2007) The therapeutic potential of statins in neurological disorders. *Curr Med Chem* 14: 103–112
 151. Wang Q, Yan J, Chen X, Li J, Yang Y, Weng J, Deng C, Yenari MA (2011) Statins: multiple neuroprotective mechanisms in neurodegenerative diseases. *Exp Neurol* 230: 27–34
 152. Tsaousidou MK, Ouahchi K, Warner TT, Yang Y, Simpson MA, Laing NG, Wilkinson PA, Madrid RE, Patel H, Hentati F *et al* (2008) Sequence alterations within CYP7B1 implicate defective cholesterol homeostasis in motor-neuron degeneration. *Am J Hum Genet* 82: 510–515
 153. Stiles AR, McDonald JG, Bauman DR, Russell DW (2009) CYP7B1: one cytochrome P450, two human genetic diseases, and multiple physiological functions. *J Biol Chem* 284: 28485–28489
 154. Schüle R, Siddique T, Deng HX, Yang Y, Donkervoort S, Hansson M, Madrid RE, Siddique N, Schöls L, Björkhem I (2010) Marked accumulation of 27-hydroxycholesterol in SPG5 patients with hereditary spastic paresis. *J Lipid Res* 51: 819–823
 155. Pitas RE, Boyles JK, Lee SH, Foss D, Mahley RW (1987) Astrocytes synthesize apolipoprotein E and metabolize apolipoprotein E-containing lipoproteins. *Biochim Biophys Acta* 917: 148–161
 156. Pitas RE, Boyles JK, Lee SH, Hui D, Weisgraber KH (1987) Lipoproteins and their receptors in the central nervous system. Characterization of the lipoproteins in cerebrospinal fluid and identification of apolipoprotein B, E (LDL) receptors in the brain. *J Biol Chem* 262: 14352–14360
 157. Xie C, Lund EG, Turley SD, Russell DW, Dietschy JM (2003) Quantitation of two pathways for cholesterol excretion from the brain in normal mice and mice with neurodegeneration. *J Lipid Res* 44: 1780–1789
 158. Lund EG, Guileyardo JM, Russell DW (1999) cDNA cloning of cholesterol 24-hydroxylase, a mediator of cholesterol homeostasis in the brain. *Proc Natl Acad Sci USA* 96: 7238–7243
 159. Karu K, Hornshaw M, Woffendin G, Bodin K, Hamberg M, Alvelius G, Sjoval J, Turton J, Wang Y, Griffiths WJ (2007) Liquid chromatography–mass spectrometry utilizing multi-stage fragmentation for the identification of oxysterols. *J Lipid Res* 48: 976–987
 160. Kotti TJ, Ramirez DM, Pfeiffer BE, Huber KM, Russell DW (2006) Brain cholesterol turnover required for geranylgeraniol production and learning in mice. *Proc Natl Acad Sci USA* 103: 3869–3874
 161. Pfeiffer S, Aivazian D (2004) Targeting Rab GTPases to distinct membrane compartments. *Nat Rev Mol Cell Biol* 11: 886–896
 162. Maioli S, Bävner A, Ali Z, Heverin M, Ismail MA, Puerta E, Olin M, Saeed A, Shafaati M, Parini P *et al* (2013) Is it possible to improve memory function by upregulation of the cholesterol 24S-hydroxylase (CYP46A1) in the brain? *PLoS ONE* 8: e68534
 163. Hudry E, Van Dam D, Kulik W, De Deyn PP, Stet FS, Ahouansou O, Benraiss A, Delacourte A, Bougnères P, Aubourg P *et al* (2010) Adeno-associated virus gene therapy with cholesterol 24-hydroxylase reduces the amyloid pathology before or after the onset of amyloid plaques in mouse models of Alzheimer's disease. *Mol Ther* 18: 44–53
 164. Lehmann JM, Kliewer SA, Moore LB, Smith-Oliver TA, Oliver BB, Su JL, Sundseth SS, Winegar DA, Blanchard DE, Spencer TA *et al* (1997) Activation of the nuclear receptor LXR by oxysterols defines a new hormone response pathway. *J Biol Chem* 272: 3137–3140
 165. Rebeck GW (2004) Cholesterol efflux as a critical component of Alzheimer's disease pathogenesis. *J Mol Neurosci* 23: 219–224

166. Tall AR (2008) Cholesterol efflux pathways and other potential mechanisms involved in the athero-protective effect of high density lipoproteins. *J Intern Med* 263: 256–273
167. Fukumoto H, Deng A, Irizarry MC, Fitzgerald ML, Rebeck GW (2002) Induction of the cholesterol transporter ABCA1 in central nervous system cells by liver X receptor agonists increases secreted Abeta levels. *J Biol Chem* 277: 48508–48513
168. Liang Y, Lin S, Beyer TP, Zhang Y, Wu X, Bales KR, DeMattos RB, May PC, Li SD, Jiang XC et al (2004) A liver X receptor and retinoid X receptor heterodimer mediates apolipoprotein E expression, secretion and cholesterol homeostasis in astrocytes. *J Neurochem* 88: 623–634
169. Matsuda A, Nagao K, Matsuo M, Kioka N, Ueda K (2013) 24(S)-hydroxycholesterol is actively eliminated from neuronal cells by ABCA1. *J Neurochem* 126: 93–101
170. Kolsch H, Lutjohann D, Tulke A, Björkhem I, Rao ML (1999) The neurotoxic effect of 24-hydroxycholesterol on SH-SY5Y human neuroblastoma cells. *Brain Res* 818: 171–175
171. Karasinska JM, Rinninger F, Lütjohann D, Ruddle P, Franciosi S, Kruit JK, Singaraja R, Hirsch-Reinshagen V, Fan J, Brunham LR (2009) Specific loss of brain ABCA1 increases brain cholesterol uptake and influences neuronal structure and function. *J Neurosci* 29: 3579–3589
172. Karasinska JM, de Haan W, Franciosi S, Ruddle P, Fan J, Kruit JK, Stukas S, Lütjohann D, Gutmann DH, Wellington CL et al (2013) ABCA1 influences neuroinflammation and neuronal death. *Neurobiol Dis* 54: 445–455
173. Wang L, Schuster GU, Hultenby K, Zhang Q, Andersson S, Gustafsson JA (2002) Liver X receptors in the central nervous system: from lipid homeostasis to neuronal degeneration. *Proc Natl Acad Sci USA* 99: 13878–13883
174. Gosselet F, Saint-Pol J, Fenart L (2014) Effects of oxysterols on the blood–brain barrier: implications for Alzheimer's disease. *Biochem Biophys Res Commun* 446: 687–691
175. Schweinzer C, Kober A, Lang I, Etschmaier K, Scholler M, Kresse A, Sattler W, Panzenboeck U (2011) Processing of endogenous AbetaPP in blood–brain barrier endothelial cells is modulated by liver-X receptor agonists and altered cellular cholesterol homeostasis. *J Alzheimers Dis* 27: 341–360
176. Ehehalt R, Keller P, Haass C, Thiele C, Simons K (2003) Amyloidogenic processing of the Alzheimer beta-amyloid precursor protein depends on lipid rafts. *J Cell Biol* 160: 113–123
177. Heverin M, Meaney S, Lütjohann D, Diczfalusy U, Wahren J, Björkhem I (2005) Crossing the barrier: net flux of 27-hydroxycholesterol into the human brain. *J Lipid Res* 46: 1047–1052
178. Mateos L, Ismail MA, Gil-Bea FJ, Schüle R, Schöls L, Heverin M, Folkesson R, Björkhem I, Cedazo-Minguez A (2011) Side chain-oxidized oxysterols regulate the brain renin-angiotensin system through a liver X receptor-dependent mechanism. *J Biol Chem* 286: 25574–25585
179. von Bohlen und Halbach O, Albrecht D (2006) The CNS renin-angiotensin system. *Cell Tissue Res* 326: 599–616
180. Pfrieger FW (2003) Role of cholesterol in synapse formation and function. *Biochim Biophys Acta* 1610: 271–280
181. Lentz BR, Carpenter TJ, Alford DR (1987) Spontaneous fusion of phosphatidylcholine small unilamellar vesicles in the fluid phase. *Biochemistry* 26: 5389–5397
182. Jahn R, Sudhof TC (1999) Membrane fusion and exocytosis. *Annu Rev Biochem* 68: 863–911
183. Chernomordik LV, Kozlov MM (2003) Protein-lipid interplay in fusion and fission of biological membranes. *Annu Rev Biochem* 72: 175–207
184. Chen Z, Rand RP (1997) The influence of cholesterol on phospholipid membrane curvature and bending elasticity. *Biophys J* 73: 267–276
185. Coorsen JR, Rand RP (1990) Effects of cholesterol on the structural transitions induced by diacylglycerol in phosphatidylcholine and phosphatidylethanolamine bilayer systems. *Biochem Cell Biol* 68: 65–69
186. Churchward MA, Rogasevskaia T, Hofgen J, Bau J, Coorsen JR (2005) Cholesterol facilitates the native mechanism of Ca²⁺-triggered membrane fusion. *J Cell Sci* 118: 4833–4848
187. Thiele C, Hannah MJ, Fahrenholz F, Huttner WB (2000) Cholesterol binds to synaptophysin and is required for biogenesis of synaptic vesicles. *Nat Cell Biol* 2: 42–49
188. Linetti A, Fratangeli A, Taverna E, Valnegri P, Francolini M, Cappello V, Matteoli M, Passafaro M, Rosa P (2010) Cholesterol reduction impairs exocytosis of synaptic vesicles. *J Cell Sci* 123: 595–605
189. Chamberlain LH, Burgoyne RD, Gould GW (2001) SNARE proteins are highly enriched in lipid rafts in PC12 cells: implications for the spatial control of exocytosis. *Proc Natl Acad Sci USA* 98: 5619–5624
190. Kudinov AR, Kudinova NV, Berezov TT (2006) Cholesterol is an important molecule in the processes of the synaptic plasticity and degeneration of neurons. *Vestn Ross Akad Med Nauk* 2006: 61–66
191. Mauch DH, Nägler K, Schumacher S, Göritz C, Müller EC, Otto A, Pfrieger FW (2001) CNS synaptogenesis promoted by glia-derived cholesterol. *Science* 294: 1354–1357
192. Göritz C, Mauch DH, Pfrieger FW (2005) Multiple mechanisms mediate cholesterol-induced synaptogenesis in a CNS neuron. *Mol Cell Neurosci* 29: 190–201
193. Lang T, Bruns D, Wenzel D, Riedel D, Holroyd P, Thiele C, Jahn R (2001) SNAREs are concentrated in cholesterol-dependent clusters that define docking and fusion sites for exocytosis. *EMBO J* 20: 2202–2213
194. Chamberlain LH, Gould GW (2002) The vesicle- and target-SNARE proteins that mediate Glut4 vesicle fusion are localized in detergent-insoluble lipid rafts present on distinct intracellular membranes. *J Biol Chem* 277: 49750–49754
195. Dason JS, Smith AJ, Marin L, Charlton MP (2014) Cholesterol and F-actin are required for clustering of recycling synaptic vesicle proteins in the presynaptic plasma membrane. *J Physiol* 592: 621–633
196. Bliss TV, Collingridge GL (1993) A synaptic model of memory: long-term potentiation in the hippocampus. *Nature* 361: 31–39
197. Malinow R, Malenka RC (2002) AMPA receptor trafficking and synaptic plasticity. *Annu Rev Neurosci* 25: 103–126
198. Malenka RC (2003) Synaptic plasticity and AMPA receptor trafficking. *Ann N Y Acad Sci* 1003: 1–11
199. Plant K, Pelkey KA, Bortolotto ZA, Morita D, Terashima A, McBain CJ, Collingridge GL, Isaac JT (2006) Transient incorporation of native GluR2-lacking AMPA receptors during hippocampal long-term potentiation. *Nat Neurosci* 9: 602–604
200. Choquet D, Triller A (2003) The role of receptor diffusion in the organization of the postsynaptic membrane. *Nat Rev Neurosci* 4: 251–265
201. Bard L, Groc L (2011) Glutamate receptor dynamics and protein interaction: lessons from the NMDA receptor. *Mol Cell Neurosci* 48: 298–307
202. Sheng M, Hoogenraad CC (2007) The postsynaptic architecture of excitatory synapses: a more quantitative view. *Annu Rev Biochem* 76: 823–847

203. Renner M, Choquet D, Triller A (2009) Control of the postsynaptic membrane viscosity. *J Neurosci* 29: 2926–2937
204. Hering H, Lin CC, Sheng M (2003) Lipid rafts in the maintenance of synapses, dendritic spines, and surface AMPA receptor stability. *J Neurosci* 23: 3262–3271
205. Martín MG, Ahmed T, Korovaichuk A, Venero C, Menchón SA, Salas I, Munck S, Herreras O, Balschun D, Dotti CG (2014) Constitutive hippocampal cholesterol loss underlies poor cognition in old rodents. *EMBO Mol Med* 6: 902–917
206. Frank C, Giammarioli AM, Pepponi R, Fiorentini C, Rufini S (2004) Cholesterol perturbing agents inhibit NMDA-dependent calcium influx in rat hippocampal primary culture. *FEBS Lett* 566: 25–29
207. Frank C, Rufini S, Tancredi V, Forcina R, Grossi D, D'Arcangelo G (2008) Cholesterol depletion inhibits synaptic transmission and synaptic plasticity in rat hippocampus. *Exp Neurol* 212: 407–414
208. Paul SM, Doherty JJ, Robichaud AJ, Belfort GM, Chow BY, Hammond RS, Crawford DC, Linsenbardt AJ, Shu HJ, Izumi Y *et al* (2013) The major brain cholesterol metabolite 24(S)-hydroxycholesterol is a potent allosteric modulator of N-methyl-D-aspartate receptors. *J Neurosci* 33: 17290–17300
209. Trovò L, Ahmed T, Callaerts-Vegh Z, Buzzi A, Bagni C, Chuah M, Vandendriessche T, D'Hooge R, Balschun D, Dotti CG (2013) Low hippocampal PI(4,5)P(2) contributes to reduced cognition in old mice as a result of loss of MARCKS. *Nat Neurosci* 16: 449–455
210. Iannilli F, Sodero AO, Ledesma MD, Dotti CG (2011) Oxidative stress activates the pro-survival TrkA pathway through membrane cholesterol loss. *Neurobiol Aging* 32: 1033–1042
211. Martín MG, Trovò L, Perga S, Sadowska A, Rasola A, Chiara F, Dotti CG (2011) Cyp46-mediated cholesterol loss promotes survival in stressed hippocampal neurons. *Neurobiol Aging* 32: 933–943
212. Liu B, Li H, Repa JJ, Turley SD, Dietschy JM (2008) Genetic variations and treatments that affect the lifespan of the NPC1 mouse. *J Lipid Res* 49: 663–669
213. Davidson CD, Ali NF, Micsenyi MC, Stephney G, Renault S, Dobrenis K, Ory DS, Vanier MT, Walkley SU (2009) Chronic cyclodextrin treatment of murine Niemann-Pick C disease ameliorates neuronal cholesterol and glycosphingolipid storage and disease progression. *PLoS ONE* 4: e6951
214. Ottinger EA, Kao ML, Carrillo-Carrasco N, Yanjanin N, Shankar RK, Janssen M, Brewster M, Scott I, Xu X, Craddock J *et al* (2014) Collaborative development of 2-hydroxypropyl- β -cyclodextrin for the treatment of Niemann-Pick type C1 disease. *Curr Top Med Chem* 14: 330–339
215. Yao J, Ho D, Calingasan NY, Pipalia NH, Lin MT, Beal MF (2012) Neuroprotection by cyclodextrin in cell and mouse models of Alzheimer disease. *J Exp Med* 209: 2501–2513
216. Porter FD, Scherrer DE, Lanier MH, Langmade SJ, Molugu V, Gale SE, Olzeski D, Sidhu R, Dietzen DJ, Fu R *et al* (2010) Cholesterol oxidation products are sensitive and specific blood-based biomarkers for Niemann-Pick C1 disease. *Sci Transl Med* 2: 56–81
217. Jiang X, Sidhu R, Porter FD, Yanjanin NM, Speak AO, te Vrugte DT, Platt FM, Fujiwara H, Scherrer DE, Zhang J *et al* (2011) A sensitive and specific LC-MS/MS method for rapid diagnosis of Niemann-Pick C1 disease from human plasma. *J Lipid Res* 52: 1435–1445
218. Sodhi RK, Singh N (2014) Liver X receptor activation by the agonist T0901317 reduces neuropathological changes and improves memory in mouse models of experimental dementia. *Eur J Pharmacol* 732: 50–59
219. Donkin JJ, Stukas S, Hirsch-Reinshagen V, Namjoshi D, Wilkinson A, May S, Chan J, Fan J, Collins J, Wellington CL (2010) ATP-binding cassette transporter A1 mediates the beneficial effects of the liver X receptor agonist GW3965 on object recognition memory and amyloid burden in amyloid precursor protein/presenilin1 mice. *J Biol Chem* 285: 34144–34154
220. Fitz NF, Castranio EL, Carter AY, Kodali R, Lefterov I, Koldamova R (2014) Improvement of memory deficits and amyloid- β clearance in aged APP23 mice treated with a combination of anti-amyloid- β antibody and LXR agonist. *J Alzheimers Dis* 41: 535–549
221. Terwel D, Steffensen KR, Verghese PB, Kummer MP, Gustafsson JÅ, Holtzman DM, Heneka MT (2011) Critical role of astroglial apolipoprotein E and liver X receptor- α expression for microglial A β phagocytosis. *J Neurosci* 31: 7049–7059
222. Wolozin B, Wang SW, Li NC, Lee A, Lee TA, Kazis LE (2007) Simvastatin is associated with a reduced incidence of dementia and Parkinson's disease. *BMC Med* 5: 20
223. Cramer C, Haan MN, Galea S, Langa KM, Kalbfleisch JD (2008) Use of statins and incidence of dementia and cognitive impairment without dementia in a cohort study. *Neurology* 71: 344–350
224. Wong WB, Lin VW, Boudreau D, Devine EB (2013) Statins in the prevention of dementia and Alzheimer's disease: a meta-analysis of observational studies and an assessment of confounding. *Pharmacoevidemiol Drug Saf* 22: 345–358
225. Trompet S, van Vliet P, de Craen AJ, Jolles J, Buckley BM, Murphy MB, Ford I, Macfarlane PW, Sattar N, Packard CJ *et al* (2010) Pravastatin and cognitive function in the elderly. Results of the PROSPER study. *J Neurol* 257: 85–90
226. Wagstaff LR, Mitton MW, Arvik BM, Doraiswamy PM (2003) Statin-associated memory loss: analysis of 60 case reports and review of the literature. *Pharmacotherapy* 23: 871–880
227. Schilling JM, Cui W, Godoy JC, Risbrough VB, Niesman IR, Roth DM, Patel PM, Drummond JC, Patel HH, Zemljic-Harpe AE *et al* (2014) Long-term atorvastatin treatment leads to alterations in behavior, cognition, and hippocampal biochemistry. *Behav Brain Res* 267: 6–11
228. Shepardson NE, Shankar GM, Selkoe DJ (2011) Cholesterol level and statin use in Alzheimer disease: II. Review of human trials and recommendations. *Arch Neurol* 68: 1385–1392
229. Chu LW, Chen JY, Yu KL, Cheng KI, Wu PC, Wu BN (2012) Neuroprotective and anti-inflammatory activities of atorvastatin in a rat chronic constriction injury model. *Int J Immunopathol Pharmacol* 25: 219–230
230. Laufs U, Gertz K, Dirnagl U, Böhm M, Nickenig G, Endres M (2002) Rosuvastatin, a new HMG-CoA reductase inhibitor, upregulates endothelial nitric oxide synthase and protects from ischemic stroke in mice. *Brain Res* 942: 23–30
231. Puttananjai MK, Dhale MA, Gaonkar V, Keni S (2011) Statins: 3-hydroxy-3-methylglutaryl-CoA (HMG-CoA) reductase inhibitors demonstrate anti-atherosclerotic character due to their antioxidant capacity. *Appl Biochem Biotechnol* 163: 215–222
232. Vance JE (2012) Dysregulation of cholesterol balance in the brain: contribution to neurodegenerative diseases. *Dis Model Mech* 5: 746–755